Appendix A Sampling and Analysis Plan Tables

Appendix A

Sampling and Analysis Plan Tables Sampling and Analysis Plan Table for Chemical and Radiological Analysis

Plan Table Number: LTS_ECM_2005

Sampling and Analysis Plan Table for Chemical and Radiological Analysis

DRAFT

Plan Table Number: LTS_ECM_2005

SAP Number: Sampler: Haney, T. J.

Date: 04/18/2005 Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING Project Manager: HANEY, T. J./VANHORN, R. L. SMO Contact: MCGRIFF, T. W.

Sampling Activity	Sample						Sample	Location		\vdash			$\overline{}$		_	$\overline{}$		$\overline{}$	\neg		\neg		_		_	_	_	_
	Type	Sample Matrix	Coll Type	Sampling Method	Planned Date	Area	Type of Location	Location	Depth (ft)	AT1		AT3	+	+	AT6 AT	+	+	+			T12 A	T13 AT1	4 AT15	AT16	AT17 A	AT18 A	T19AT	20
ECR141	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	DEER MOUSE	REFERENCE AREA	NA NA				+	5	1	1	+	1	1	+	1	$^{+}$	$^{+}$	Н	†	$^{+}$	†	-
ECR142	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	DEER MOUSE	REFERENCE AREA	NA NA		П		\exists	5	١,	1		1	1	1	1		T	П	T	\top	T	-
ECR143	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	DEER MOUSE	REFERENCE AREA	NA NA		П		T	5	1	1	1	1	1	1	1		Т	П	T	寸	\top	_
ECR144	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	DEER MOUSE	REFERENCE AREA	NA		П		T	5	1	1	1	1	1	1	1		T	П	T	\neg	\top	
ECR145	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	DEER MOUSE	REFERENCE AREA	NA		П		T	5	1	1	1	1	1		1	T	П	П	1	T	T	1
ECR146	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	DEER MOUSE	REFERENCE AREA	NA				T	5	1	1		1	1		1		T	П	T	T	T	٦
ECR147	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	DEER MOUSE	REFERENCE AREA	NA		П		П	5	1	1		1	1		1		П	П	T	T	T	
ECR148	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	DEER MOUSE	REFERENCE AREA	NA		П		T	5	1	1		1	1		1		Т	П	T	1	\top	
ECR149	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	DEER MOUSE	REFERENCE AREA	NA		П		\exists	5	1	1	1	1	1		1		T	П	\forall	\top	\top	7
ECR150	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	DEER MOUSE	REFERENCE AREA	NA		П		T	5	1	1		1	1		1		T	П	T	\exists	\top	Ī
ECR151	REG/QC	PLANT BIOTA	DUP	COMP	06/01/2005	INEEL	SAGEBRUSH	REFERENCE AREA	NA		П		\exists	T	- 2	2		2	2		2		T	П	T	\neg	\top	Ī
ECR152	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	REFERENCE AREA	NA		П		\exists	T	1	1		1	1		1	T	Т	П	T	\top	\top	Ī
ECR153	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	REFERENCE AREA	NA		П		\exists	T	1	1		1	1		1		T	П		\top	\top	7
ECR154	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	REFERENCE AREA	NA		П		\exists		1	1		1	1	1	1		Т	П		\top	\top	Ī
ECR155	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	REFERENCE AREA	NA		П		\exists		1	1		1	1	1	1	T	T	П	1	1		7
he sampling activ	vity displayed o	n this table represents th	e first 6 to 9	characters	of the sample ide	ntification number.	he complete sample identification	number will appear on the sample I	labels.						D-D	louble	QC V	olum	• 1	T - Trij	ple Q	Volum	0					_
T1: Analysis Su	Suite #1						T11: Radiochemistry - Suite 2						Comme		Ordnan	ne Di	enneal	Area										
T2: Analysis St	uite #2						Total Metals (TAL)					-	_		etonatio			Alta									=	
T3: Analysis Su	Suite #3						XT13:								Explosio		a											
T4: Gamma Sc	creen						T14:							_	a North	_											_	
T5: Histapthy							XT15:						BLRS: E	Big Los	t River S	Sinks											_	
T6: Hydrogen I	lon (pH)						XT16:					_ 7	Inalysis	s Suite	#3 and	gamn	na scre	een ar	e field	scree	ning te	ests.					=	
T7: Mercury							XT17:					_ [_	
T8: Mesoarthop	ppod						KT18:									_			_								_	
T9: Nitroaroma	atics (8330)						XT19:																				_	
T10: Radiochem	mistry - Suite 1						T20:																				=	
Analysis Suites:								Contingencies:																				
vnalysis Suite #1.	Moisture Cont	lent, Hydrogen Ion (pH), (Cation Excha	ange Capac	ity																							
		xicity Test, Rye Grass G	rowth Test																								_	
Analysis Suite #3:																											_	
		1, Gamma Spec, Pu-Iso,).																							_	
adiochemistry - 3	Suite 2: Gross	Alpha, Gross Beta, Gami	па эрес																								-	
																											_	
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Plan Table Number: LTS_ECM_2005

SAP Number: Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

DRAFT

Sampler: Haney, T. J. Project Manager: HANEY, T. J./VANHORN, R. L. SMO Contact: MCGRIFF, T. W.

Sergis Se	Sergiely Surgiely Sur		5	Sample Description					Sample	Location		L					9	Enter /	Analysi	із Туре	s (AT)	and C	Quantity (Reques	ted			
Control Type Mark Type Mark Type Mark Control Co	Control Type	0200000000		m SE		L			376		I was	AT1	AT2	АТЗ	AT4	AT5 A	T6 AT	7 AT	8 ATS	AT1	0 AT11	1 AT12	AT13	AT14 A	T15 AT1	6 AT1	AT18	AT19
Control Record Part Biol Part Biol Record Rec	CRESTS REG	Sampling Activity			(39.5)	100000000000000000000000000000000000000	A. M. S. C.	Area		Location	120 100 100 100	за	9A	1G	R5	3Z F	н но	3 3Y	N7	RH	RN	LA	\prod	\top	\pm	\top	H	
Part Biota Graph Comp Gold Comp	SECRISS REG	ECR156	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	REFERENCE AREA	NA						1	\perp	1	1		1						
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ECRISO REG PLANT BIOTA GRAB COMP 06910205 NEEL SAGEBRUSH REFERENCE AREA NA I I I I I I I I I I I I I I I I I I	SCRIEGO PLANT BIOTA GRAD COMP GOSTIZODO NEEL SAGEBRUSH REFERENCE AREA NA	ECR158	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	REFERENCE AREA	NA						1	\blacksquare	1	1		1	П			П		
ECR161 REGOC PLANT BOTA DUP COMP 06010005 NEEL CRESTED WHEATOR REFERENCE AREA NA IN 1 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2	Control REGOOD PLANT BIOTA DUP COMP 0601/2005 NEEL CRESTED WHEATOR REFERENCE AREA NA	ECR159	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	REFERENCE AREA	NA		Г		П		1		1	1	П	1	П				П	
ECR162 REG PLANT BIOTA GRAB COMP 0601/2005 NEEL CRESTED WHEATOR REFERENCE AREA NA I I I I I I I I I I I I I I I I I I	ECR162 REG PLANT BIOTA GRAB COMP 0601/2005 NEEL CRESTED WHEATOR REFERENCE AREA NA I I I I I I I I I I I I I I I I I I	ECR160	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	REFERENCE AREA	NA	Г	Т	Г	П	\exists	1		1	1		1	П	T		\top	П	Γ
ECR143 REG PLANT BIOTA GRAB COMP 0601/2005 NEEL CRESTED WHEATGR REFERENCE AREA NA I I I I I I I I I I I I I I I I I I	CERTIES REG PLANT BIOTA GRAB COMP 0601/2005 NEEL CRESTED WHEATOR REFERENCE AREA NA 1 1 1 1 1 1 1	ECR161	REG/QC	PLANT BIOTA	DUP	COMP	06/01/2005	INEEL	CRESTED WHEATGR	REFERENCE AREA	NA	Г	Т	Г	П	\exists	2	2	2	2	\Box	2	П	\top	\top	\top	П	Г
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ECR165 REG PLANT BIOTA GRAB COMP 0601/2005 NEEL CRESTED WHEATOR REFERENCE AREA NA I I I I I I I I I I I I I I I I I I	ECR165 REG PLANT BIOTA GRAB COMP 0691/2005 NEEL CRESTED WHEATOR REFERENCE AREA NA I I I I I I I I I I I I I I I I I I	ECR163	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	REFERENCE AREA	NA	Г	Т	Г	П	\exists	1		1	1	\Box	1	П		T	\vdash	П	Г
ECR166 REG PLANT BIOTA GRAB COMP 0601/2005 NEEL CRESTED WHEATOR REFERENCE AREA NA I I I I I I I I I I I I I I I I I I	ECR166 REG PLANT BIOTA GRAB COMP 0601/2005 NEEL CRESTED WHEATGR REFERENCE AREA NA 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECR164	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	REFERENCE AREA	NA	Г	Т		П		1		1	1	Т	1	П			T	П	Г
ECR167 REG PLANT BIOTA GRAD COMP 0601/2005 NEEL CRESTED WHEATGR REFERENCE AREA NA IN	ECR167 REG PLANT BIOTA GRAB COM 0601/2005 NEEL CRESTED WHEATOR REFERENCE AREA NA I I I I I I I I I I I I I I I I I I	ECR165	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	REFERENCE AREA	NA	Г	Т		П	\top	1		1	1	\vdash	1	Ħ	\top	$^{+}$	†	П	Г
ECR168 REG PLANT BIOTA GRA8 COMP 0601/2005 INEEL CRESTED WHEATOR REFERENCE AREA NA IN	ECR168 REG PLANT BIOTA GRAB COMP 06:017:005 NEEL CRESTED WHEATOR REFERENCE AREA NA	ECR166	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	REFERENCE AREA	NA	Т	T	Т	П	\forall	1		1	1	\vdash	1	П	\top	+	†	Н	Г
ECR159 REG PLANT BIOTA GRAB COMP 6601/2005 NEEL CRESTED WHEATGR REFERENCE AREA NA I I I I I I I I I I I I I I I I I I	ECR169 REG PLANT BIOTA GRAB COMP 06/01/2005 INEEL CRESTED WHEATGR REFERENCE AREA NA 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECR167	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	REFERENCE AREA	NA	T	T	Т	Н	\top	1		1	1	\vdash	1	H	\top	+	十	Н	Г
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sampling activity displayed on this table represents the first 6 to 9 characters of the sample identification number. The complete sample identification number will appear on the sample labels. AT11: Radiochemistry: State 2 Comments: AT12: Total Metals (TAL.) MDA: Mava Bordnance Disposal Area MDA: Radiochemistry: State 9 RAIL: Ralicar Explosion area TAN: Test Area North BLRS: Big Loat River Sinks AT16: Analysis State #3 AT17: Analysis State #3 AT18: Analysis State #3 AT18: Analysis State #3 AT18: Analysis State #3 AT18: Analysis State #3 AT19: Analysis Analy	sampling activity displayed on this table represents the first 6 to 9 characters of the sample identification number. Analysis Suite #1 AT11: Radiochemistry - Suite 2 Analysis Suite #3 AT12: Total Metals (TAL) Analysis Suite #3 AT14: Total Metals (TAL) Analysis Suite #3 AT15: Total Metals (TAL) AT16: BLRS: Big Loat River Sinks BLRS: Big Loat River Sinks Analysis Suite #3 and gamma screen are field screening tests. Mercury AT16: Analysis Suite #3 AT17: Mesourthopod AT18: Merouny AT19: Mesourthopod AT18: Suite #3 AT19: Contingencies: AT19: Contingen	ECR169	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	REFERENCE AREA	NA	T	T	Т	П	\exists	1		1	1	\vdash	1	П	\top	\top	\vdash	П	Г
s ampling activity displayed on this table represents the first 6 to 9 characters of the sample identification number. The complete sample identification number will appear on the sample labels. Comments: AT11: Radiochemistry: State 2 AT22: Total Metals (TAL) MDA: Mass Belloration Area ATAN: Test Area North ATAN: Test Area North BLRS: Big Lost River Sinks AT41: Analysis State #3 AT41: Analysis State #3 AT41: Analysis State #3 AT41: Analysis Availate #3 ANAlysis Availate #4 ANAlysis Availate #3 ANAlysis Availate #4 ANAlysis A	sampling activity displayed on this table represents the first 6 to 9 characters of the sample identification number. Analysis Suite #1 AT11: Radiochemistry - Suite 2 Analysis Suite #3 AT12: Total Metals (TAL) Analysis Suite #3 AT14: Total Metals (TAL) Analysis Suite #3 AT15: Total Metals (TAL) AT16: BLRS: Big Loat River Sinks BLRS: Big Loat River Sinks Analysis Suite #3 and gamma screen are field screening tests. Mercury AT16: Analysis Suite #3 AT17: Mesourthopod AT18: Merounded (R330) Or Radiochemistry - Suite 1 AT19: Contingencies:	ECR170	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	REFERENCE AREA	NA NA	H	\vdash	\vdash	H	+	١,	+	1	1	\vdash	1	H	1	+	+	Н	М
Hydrogen Ion (pH)	Hydrogen lon (pH) AT16: Analysis Suite #3 and gamma screen are field screening tests. Mercury AT17: Mesoarthopod AT18: Nitroaromatics (8330) AT19: Radiochemistry - Suite 1 AT20: AT30: AT30: AT30: Confingencies: AT30: AT30	Analysis Analysis Analysis Gamma S	Suite #1 Suite #2 Suite #3 Screen	on this table represents the	he first 6 to 9) characters	of the sample ide	A A A	T11: Radiochemistry - Suite 2: T12: Total Metals (TAL) T13: T14:	650	labels.			_	NODA: MDA: I RAIL: I TAN: T	Naval Mass De Railcar E Test Area	Ordnano tonation explosion	ce Disp n Area on area	posal A		1.	Triple	QC Vol	ume		<u> </u>	<u> </u>	
9. Nitroaromatics (8330) AT19: 10: Radiochemistry - Suitle 1 Alysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity alysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test alysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test alysis Suite #3: Mercury, Nitroaromatics (8330) diliochemistry - Suite 1: Am-241, Gamma Spec, Pu-lso, U-lso, Si-90	Nitroaromatics (8330) O: Radiochemistry - Suite 1 A720: alysis Suite # 1: Moisture Conlent, Hydrogen Ion (pH), Cation Exchange Capacity slysis Suite # 2: Earthworm Toxicity Test, Rye Grass Growth Test slysis Suite # 2: Earthworm Toxicity Test, Rye Grass Growth Test slysis Suite # 3: Mercury, Nitroaromatics (8330) diochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90	6; Hydroger 7: Mercury	n lon (pH)					A	T16:					_ _ _	Analysi	is Suite	43 and g	jamma	a scree	en are l	ield scr	reenin	g tests.					
10: Radiochemistry - Suite 1 alysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity alysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test alysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test alysis Suite #3: Mercury, Nitroaromatics (8330) dilochemistry - Suite 1: Am-241, Gamma Spec, Pu-lso, U-lso, Si-90	0. Radiochemistry - Suite 1 A720: alyais Suite #1: Moisture Conlent, Hydrogen Ion (pH), Cation Exchange Capacity alyais Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test alyais Suite #3: Mercury, Nitroaromatics (8330) diochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90													- !	_			_			_	_		_	_	_	_	_
alysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity alysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test alysis Suite #3: Mercury, Nitroaromatics (8330) stillochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90	Alysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity Alysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Alysis Suite #3: Mercury, Nitroaromatics (8330) Side #3: Mercury, Nitroaromatics (8330)			8					-					7	_			_			_	_		_	_	_	_	_
alysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test alysis Suite #3: Mercury, Nitroaromatics (8330) stillochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90	alyais Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test alyais Suite #3: Mercury, Nitroaromatics (8330) diochemistry - Suite 1: Arn-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90	alysis Suites:							AUTHO M	Contingencies:				_														
alysis Suite #3: Mercury, Nitroaromatics (8330) ktlicohemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90	slysis Suite #3: Mercury, Nitroaromatics (8330) tiochemistry - Suite 1: Arn-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90	nalysis Suite #	1: Moisture Con	itent, Hydrogen Ion (pH),	Cation Exch	ange Capac	city																					_
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	and received 1 - outre 2. Order Aprile, Order Deal, Delining Open					U												_			_	_		_			_	_

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Plan Table Number: LTS_ECM_2005

SAP Number:

Mark		S	ample Description					Sample	Location		L					En	ter An	alysis	Types	(AT) a	ind Qua	ntity Re	quested	1		
ECRIT REGIO SOL OUR OU	Sampling			(38/3)	100000000000000000000000000000000000000	A1000000000000000000000000000000000000			W	C20 (1980) 18 / V	\vdash	Н	AT3	-	+	+	AT8	AT9	AT10	AT11	AT12 A	T13 AT	14 AT15	AT16 A	T17 A7	T18 A
ECRITY RED SOL GAME COMP GO110005 NEEL SUNFACE SOLS REFERENCE AREA 0 2 NOVES 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	Activity	Туре	Matrix	Type	Method	Date	Area	Location	Location	(ft)	3A	9A	1G	R5	3Z PH	HG	3Y	N7	RH	RN	LA				\perp	
ECRITO REG SOLI GIAMA COMP 06112005 NEEL SURFACE SOLS NEFERENCE AREA 0 2 NOHES 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	ECR171	REG/QC	SOIL	DUP	COMP	06/01/2005	INEEL	SURFACE SOILS	REFERENCE AREA	0-2 INCHES	2					2		2	2		2	-		T f		
ECRIT RED SOIL GRAD COMP 0010005 NEEL SURFACE SOLS REFERENCE AREA 0 2 NO-RES 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	ECR172	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOILS	REFERENCE AREA	0-2 INCHES	1.					1		1.	1		1.					
ECRIT REG SOIL GRAB COMP 06110005 NEEL SURFACE SOILS REFERENCE AREA 0-2 NCHES 1 0 0 0 0 0 0 0 0 0	ECR173	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOILS	REFERENCE AREA	0-2 INCHES	1	П		П		1	Г	1	1	П	1			П	Т	Т
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ECRIT 7 REG SOIL GRAB COMP 0612005 NEEL SURFACE SOILS REFERENCE AREA 0.2 NICHES 1 0 0 1 0 1 0 1 0 1 0 1 0 0 0 0 0 0 0	ECR175	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOILS	REFERENCE AREA	0-2 INCHES	1	П		\dashv	\top	1		1	1	П	1	\top	T	П	\top	\uparrow
ECR179 REG SOIL GRAB COMP 06010005 NEEL SURFACE SOILS REFERENCE AREA 0.2 NCHES 1 1 0 1 0 1 0 1 0 1 0 1 0 1 0 0 0 0 0	ECR176	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOILS	REFERENCE AREA	0-2 INCHES	1	H		\dashv	\top	1		1	1	П	1	\top	T	П	$^{+}$	†
ECR119 REG SOIL GRAB COMP 06012005 NEEL SURFACE SOLS REFERENCE AREA 0 2 NCHES 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	ECR177	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOILS	REFERENCE AREA	0-2 INCHES	1	Н	П	\dashv	+	1	T	1	1	Н	1	1	1	Н	+	†
SCIR SCIR SCIR SCIR SCIR SCIR SCIR SCIR SCIR SUBSURFACE SOIL REFERENCE AREA 0.2 INCHES 1	ECR178	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOILS	REFERENCE AREA	0-2 INCHES	1	H		\forall	\top	1		1	1	Н	1	+	T	Н	+	†
ECR181 REGIG SOIL DIP COMP 066112005 NEEL SUBSURFACE SOIL REFERNCE AREA 2-24 NCHES 2	ECR179	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOILS	REFERENCE AREA	0-2 INCHES	1	Н		\dashv		1		1	1	Н	1	+	1	H	+	†
ECR181 REGIG SOIL DIP COMP 066112005 NEEL SUBSURFACE SOIL REFERNCE AREA 2-24 NCHES 2	ECR180	REG	SOIL	GRAB	COMP	06/01/2005	INFEL	SURFACE SOILS	REFERENCE AREA	0-2 INCHES	1	H	_	\dashv	+	1		1	1	H	1	+	+	Н	+	+
ECR182 REG SOIL GRAB COMP 0691/2005 NEEL SUBSURFACE SOIL REFERENCE AREA 2.24 NCHES 1 0 0 0 1 0 1 0 1 0 0 0 0 0 0 0 0 0 0		10000	F-1370 B	00.000		201201120011	COMME	100 000 000 000 000 000 000 000 000 000	7300-03000000000000	3(m.) (20) (20)	-	Н	-	\dashv	+	2	Н	2	2	Н	2	+	+	Н	+	+
ECR183 REG SOIL GRAB COMP 0601/2005 NEEL SUBSURFACE SOIL REFERNCE AREA 2.24 INCHES 1	STREET, STREET	(0.7 C2/03/04/33)) Directo	2000	1208.000.000) DEPENDE	The thirt was an accordance to	UNIA PROGRESSION OF SEEDING	Lessen entre entre la	+	Н	_	\dashv	+	+	_		274.	Н	1	+	+	Н	+	+
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e sampling activity displayed on this table represents the first 6 to 9 characters of the sample identification number. The complete sample identification number will appear on the sample labels. D - Double QC Volume T - Triple QC Volume Comments: NOA: Navail Ordnance Disposal Area MGA: Mass Detonation Area AT11: Radiochemistry - Suite 2 Total Metals (TAL) MGA: Mass Detonation Area RAIL: Ralicar Explosion area TAN: Test Area North BLRS: Big Lost River Sinks Analysis Suite #3 AT14: Analysis Suite #3 AT15: Analysis Suite #3 A			7.50	-							+	Н	_	\dashv	+	+	_	Н	_	Н	1	-	+	Н	+	+
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Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

Project Manager: HANEY, T. J./VANHORN, R. L. SMO Contact: MCGRIFF, T. W.

Sampler: Haney, T. J.

	S	Sample Description					Sample	Location		L		_		_	En	nter An	nalysis	Туре	(AT)	and Qu	antity F	Request	ed		_	_
Sampling Activity	Sample Type	Sample Matrix	Coll Type	Sampling Method	Planned Date	Area	Type of Location	Location	Depth (ft)	AT1	Н	AT3	-	AT5 AT6	\vdash	AT8	AT9		AT11	AT12	AT13 /	T14 AT	15 AT16	AT17	AT18 A	T19A
ECR186	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SUBSURFACE SOIL	REFERENCE AREA	2-24 INCHES	1	Н	Н	\forall	+	1	Н	1	1	Н	1	\forall	1		H	\top	\dagger
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ECR188	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SUBSURFACE SOIL	REFERENCE AREA	2-24 INCHES	1	П	П	T	\top	1	Г	1	1	П	1	T	1	T	П	\top	7
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ECR193	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SOIL	REFERENCE AREA	0-3 INCHES	T	H		\forall		Н	1	Н		Н		Ħ	\top	T	H	\top	7
ECR194	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SOIL	REFERENCE AREA	0-3 INCHES	T	Ħ		\dashv	1	Т	1	H	Н	H		\forall	1	T	H	\top	
ECR195	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SOIL	REFERENCE AREA	0-3 INCHES	\vdash	Н	H	\dashv		Н	1	Н		Н	\dashv	\dashv	+	+	H	\dashv	=
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ECR197	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SOIL	REFERENCE AREA	0-3 INCHES	\vdash	Н	Н	\dashv	+	\vdash	1	\vdash	\vdash	Н	\dashv	\dashv	+	+	H	+	_
ECR198	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SOIL	REFERENCE AREA	0-3 INCHES	\vdash	Н	Н	\dashv	+	Н	1	H	H	Н	\dashv	\dashv	+	+	H	+	-
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2: Analysis	Suite #2						AT12: Total Metals (TAL)					_	_	Naval On		_	sal Are	ea								_
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T5: Histapthy						-	AT15:					- 1	BLRS: E	Big Lost R	iver Sini	ks										
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T9: Nitroaror	natics (8330)					-	AT19:					٠						_								_
T10: Radioch	emistry - Suite 1	3					AT20:					_ :														
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Plan Table Number: LTS_ECM_2005

SAP Number: Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

Project Manager: HANEY, T. J./VANHORN, R. L.

Sumple S				0.0		ING TERM ECO				Project Manager: HANE1	.,																		
Serving Surving Surving Marks Surving Marks		S	Sample Description						Sample	Location								Enter	Analy:	is Typ	es (Al	r) and	Quan	ity Req	uested				
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ECR211 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR212 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR213 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR214 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR215 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR216 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR217 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR218 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTAL REFERENCE AREA MACKAY RES ECR219 REG PLANT BIOTA COMP GRAB 0601/2005 INEEL CATTA	ECR209	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	T	SOIL	REFERENCE AREA	0-12 INCHES	Г	1		П	П				T	T	T	T				1	1	_
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ECR215 REG PLANT BIOTA COMP GRAB 66 01/2005 NEEL CATTAL REFERENCE AREA MACKAY RES To Analysia Sulte #1 Analysia Sulte #1 Analysia Sulte #2 Analysia Sulte #2 Analysia Sulte #3 Comments: NOOA: Naval Cortannoe Disposal Area NOOA: Nav	ECR213	REG	PLANT BIOTA	COMP	GRAB	06/01/2005	INEEL	T	CATTAIL	REFERENCE AREA	MACKAY RES	Г			П					1		1						T	
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AT15: Histapthy AT16: Hydrogen lon (pH) AT16: Analysis Quite #3 and gamma screen are field screening tests. AT17: Mercury AT17: Mescarthopod AT18: Analysis Suite #3 and gamma screen are field screening tests. AT18: Mescarthopod AT18: AT18	73: Analysis	s Suite #3						AT13:											а										_
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SAP Number:

Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

Project Manager: HANEY, T. J./VANHORN, R. L.

Servicing Serv	Semple S		5	Sample Description						Sample	e Location									Enter /	Analysi	в Туре	es (AT)	and Q	antity F	(eques	ted	_		_
ECR214 REG	ECR216 RG			S2500 117 P	(36.5)		1100000000	Area	T		Location	n	12000	_			\dashv	+	+	-	+	+	-		AT13 A	T14 A	T15 AT	16 AT1	7 AT1	8 AT19A
EOCI219 REG	ECR219 REG WATER GRAB COMP 0601005 NEEL POND WATER REFERENCE AREA MACKAY RES 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECR216	REG	WATER	GRAB	COMP	06/01/2005	INEEL		POND WATER	REFERENCE	E AREA	MACKAY RES	H	H	H	\forall	\forall		+	t	1		1		†		t	t	H
ECR219 REG	ECR219 REG WATER ORAB COMP 00:10055 NEEL POND WATER REFERENCE AREA MACKAY RES 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECR217	REG	WATER	GRAB	COMP	06/01/2005	INEEL		POND WATER	REFERENCE	E AREA	MACKAY RES		\Box			T		t	T	1		1:	\top		1	t	T	\Box
ECR221 REG	ECR221 REG	ECR218	REG	WATER	GRAB	COMP	06/01/2005	INEEL	1	POND WATER	REFERENCE	EAREA	MACKAY RES	Г	T		T	T	- 19		T	1		1	\exists	1	Ť	T	T	П
ECR221 REG ANNAL BOTA GRAB COMP 06012005 NEEL TADPOLEFROG REFERENCE AREA MACKAY RES	ECR221 REG	ECR219	REG	WATER	GRAB	COMP	06/01/2005	INEEL	1	POND WATER	REFERENCE	EAREA	MACKAY RES	Г	П		T	T				1	П	1	\exists			T	Т	П
ECR222 REG ANIMAL BIDTA GRAB COMP 601/2005 NEEL TADPOLEFROO REFERENCE AREA MACKAY RES	ECR222 REG	ECR220	REG	WATER	GRAB	COMP	06/01/2005	INEEL		POND WATER	REFERENCE	E AREA	MACKAY RES				\exists	T				1		1	\Box	T		T	T	\Box
ECR223 REG	ECR223 REG ANIMAL BIOTA GRAB COMP 0601/2005 NEEL TADPOLEFROQ REFERENCE AREA MACKAY RES	ECR221	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	T	TADPOLE/FROG	REFERENCE	EAREA	MACKAY RES	П			\exists	T		T	Т	1		1	\exists	T		T	T	\Box
ECR224 REG	ECR224 REG ANIMAL BIOTA GRAB COMP 661/2005 NEEL TADPOLEFROG REFERENCE AREA MACKAY RES 1 1 1	ECR222	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	T	TADPOLE/FROG	REFERENCE	E AREA	MACKAY RES				П	T				1		1	П	T		T	T	П
ECR225 REG ANIAM BIOTA GRAB COMP 06/01/2005 NEEL SEDMENT REFERENCE AREA MACKAY RES 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECR225 REG ANIMAL BIOTA GRAB COMP 06012005 NEEL TADPOLEFROG REFERNCE AREA MACKAY RES	ECR223	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	1	TADPOLE/FROG	REFERENCE	E AREA	MACKAY RES					\Box		T		1		1	T				Т	П
ECR226 REG SEDMENT GRAB COMP 0601/2005 NEEL SEDMENT REFERENCE AREA MACKAY RES 1 1 1 1 1 1 1 1 1	ECR226 REG SEDMENT GRAB COMP 0601/2005 NEEL SEDMENT REFERENCE AREA MACKAY RES 1 1 1 1 1 1 1 1 1	ECR224	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	1	TADPOLE/FROG	REFERENCE	E AREA	MACKAY RES	П			T	T				1		1.	T					П
ECR227 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR228 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR229 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACKAY RES ECR230 REG SEDMENT GRAB COMP 06012005 INEEL SEDMENT REFERENCE AREA MACK	ECR227 REG SEDMENT GRAB COMP 0601/2005 NEEL SEDMENT REFERENCE AREA MACKAY RES 1 1 1 1 1 1 1 1 1	ECR225	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	T	TADPOLE/FROG	REFERENCE	EAREA	MACKAY RES	Г			\exists	T			T	1		1				T	T	П
ECR228 REG SEDMENT GRAB COMP 0601/2005 NEEL SEDMENT REFERENCE AREA MACKAY RES 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECR228 REG	ECR226	REG	SEDIMENT	GRAB	COMP	06/01/2005	INEEL	1	SEDIMENT	REFERENCE	EAREA	MACKAY RES				\exists	\exists	1			1		1		T	1	T	T	П
ECR229 REG SEDIMENT GRAB COMP 0601/2005 NEEL SEDIMENT REFERENCE AREA MACKAY RES 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECR229 REG SEDIMENT GRAB COMP 0601/2005 NEEL SEDIMENT REFERENCE AREA MACKAY RES ECR230 REG SEDIMENT GRAB COMP 0601/2005 NEEL SEDIMENT REFERENCE AREA MACKAY RES The sampling activity displayed on this table represents the first 6 to 9 characters of the sample identification number. ATI: Analysis Suite #1 ATI: Analysis Suite #2 ATI: Analysis Suite #2 ATI: Analysis Suite #3 Analysis Suite #4 Analysis	ECR227	REG	SEDIMENT	GRAB	COMP	06/01/2005	INEEL	1	SEDIMENT	REFERENCE	E AREA	MACKAY RES	Г	П	\exists	T	T	1	T	T	1		1	\exists	T		T	T	П
ECR230 REG SEDIMENT GRAB COMP 06/01/2005 NEEL SEDIMENT REFERENCE AREA MACKAY RES 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECR230 REG SEDMENT GRAB COMP 06/01/2005 NEEL SEDMENT REFERENCE AREA MACKAY RES 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECR228	REG	SEDIMENT	GRAB	COMP	06/01/2005	INEEL	1	SEDIMENT	REFERENCE	E AREA	MACKAY RES	Г			\exists	T	1		T	1		1		T		T	T	П
The sampling activity displayed on this table represents the first 6 to 9 characters of the sample identification number. AT1: Analysis Suite #1 AT1: Radiochemistry - Suite 2 AT1: Analysis Suite #2 AT1: Analysis Suite #3 AT2: Total Metals (TAL) AT3: Analysis Suite #3 AT4: Gamma Screen AT1: Analysis Suite #3 AT4: Gamma Screen AT1: History AT1: History AT1: History AT1: History AT1: History AT1: History AT1: Analysis Suite #3 AT1: History AT1:	The sampling activity displayed on this table represents the first 6 to 9 characters of the sample identification number. AT1: Analysis Suite #1 AT2: Analysis Suite #2 AT3: Analysis Suite #3 AT4: Gamma Screen AT4: Gamma Screen AT5: Histopthy AT6: Hydrogen lon (pH) AT7: Mercury AT7: New Contended (8330) AT19: Analysis Suite #3 AT19: Marcaromatics (8330) AT19: Analysis Suite #1 AT20: Contingencies: Contingencies: Contingencies: Contingencies: Analysis Suite #3 Entrucy, Viscorantics (8330) Radiochemistry - Suite 1: Analysis Content, Hydrogen lon (pH), Cation Exchange Capacity Analysis Suite #3 Entrucy, Viscorantics (8330) Radiochemistry - Suite #3 Mercury, Viscorantics (8330) Radiochemistry - Suite #4 Mercury - Mercury (84) Radiochemistry - Suite #4 Mercury - Mercury (84) Radiochemistry - Suite #4 Mercury - Mercury (84) Rad	ECR229	REG	SEDIMENT	GRAB	COMP	06/01/2005	INEEL		SEDIMENT	REFERENCE	E AREA	MACKAY RES	П	П		П	T	1		Т	1	П	1	П				Τ	П
AT1: Analysis Sulte #1	AT1: Analysis Sulte #1	ECR230	REG	SEDIMENT	GRAB	COMP	06/01/2005	INEEL		SEDIMENT	REFERENCE	E AREA	MACKAY RES	П	П		T	T	1		Т	1	П	1	П	1		Т	Т	П
AT13: Analysis Sulte #3 AT14: AT15:	AT3: Analysis Suite #3	T1: Analysis !	Suite #1	on this table represents th	ne first 6 to 9	characters	of the sample ide	entification number.	AT11:	Radiochemistry - Suite 2	. 1000	n the sample lat	bels.			- <u>r</u>	NODA:	Naval	Ordnan	ce Dis	posal A		1-1	Triple	QC Volu	ime				
AT5: Histapthy AT6: Hydrogen lon (pH) AT6: Hydrogen lon (pH) AT7: Mercury AT7: Mercury AT8: Mesoarthopod AT8: Mesoarthopod AT9: AT9: AT9: Nitroaromatics (830) AT9: AT8: Mesoarthopod AT9: AT9: AT8: Mesoarthopod AT9: AT9: AT8: Mesoarthopod AT9: AT9: AT9: Nitroaromatics (830) AT9: AT9: Nitroaromatics (830) AT9: AT9: Nitroaromatics (830) AT9: Analysis Suites: Confingencies: Confingencies: Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-lso, U-lso, St-90 Rediochemistry - Suite 1: Am-241, Gamma Spec, Pu-lso, U-lso, St-90	ATS: Histapthy AT15:	T3: Analysis	Suite #3						AT13:							-														
AT5: Hstaghty AT15: Hydrogen lon (pH) AT16: Analysis Suite #3 and gamma screen are field screening tests. AT7: Mercury AT8: Mesoarthopod AT17: AT18: Mesoarthopod AT18: AT19: AT19: Nitroaromatics (8330) AT19: AT10: Radiochemistry - Suite 1 AT20: Analysis Suite #3 and gamma screen are field screening tests. AT9: Nitroaromatics (8330) AT19: Confingencies: Confingencies: Analysis Suite #3. Mercury, Officencies (Confingencies: Analysis Suite #3. Mercury, Nitroaromatics (8330) Analysis Suite #3. Mercury, Nitroaromatics (8330) Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-lso, U-lso, Sr-90 Rediochemistry - Suite 1: Am-241, Gamma Spec, Pu-lso, U-lso, Sr-90	ATS: Histapthy AT16:	T4: Gamma S	Screen						AT14:																					
AT1: Mercury AT1: Mercury AT8: Mesoarthopod AT18:	AT7: Mercury AT18: Mesoarthopod AT18:	T5: Histapthy							AT15:							_ =	BLRS: E	Sig Los	t River	Sinks		_		_						
AT8: Mesoarthopod AT18: AT9: Ntroarcomatics (8330) AT19: AT10: Radiochemistry - Suite 1 AT20: Confingencies: Analysis Suite #1: Moisture Conlent, Hydrogen Ion (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Ntroarcomatics (8330) Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90	AT8: Mesoarthopod AT18:	T6: Hydrogen	n Ion (pH)							/						_ 7	Analysis	Suite	#3 and	gamma	a scree	n are	field scr	eening	tests.					=
AT9: Nitroaromatics (8330) AT10: Radiochemistry - Suite 1 Aralysis Suites: Analysis Suite #1: Moisture Conlent, Hydrogen Ion (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroaromatics (8330) Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90	AT19: Nitroaromatics (8330) AT19: AT20: Analysis Suites 1 Analysis Suite #1: Moisture Conlent, Hydrogen Ion (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroaromatics (8330) Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90									-								_				_		_						
AT10: Radiochemistry - Suite 1 Analysis Suites: Analysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroarcmatics (8330) Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90	AT10: Radiochemistry - Suite 1 Analysis Suites: Analysis Suite #1: Moisture Conlent, Hydrogen Ion (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroaromatics (8330) Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90									·						-]														
Analysis Suites: Analysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroaromatics (8330) Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90	Analysis Suites: Analysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroaromatics (8330) Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90			i i																										_
Analysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroaromatics (8330) Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90	Analysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroaromatics (8330) Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90	1000	unio i						AIZU.	-	Continge	encies:																		_
Analysis Suite #3: Mercury, Nitroaromatics (8330) Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-lso, U-lso, Sr-90	Analysis Suite #3: Mercury, Nitroaromatics (8330) Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90	and the state of the	1: Moisture Con	ntent, Hydrogen Ion (pH).	Cation Exch	ange Capac	ity																							
Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-lso, U-lso, Sr-90	Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90				Frowth Test																									
radiocnemosty - suite Z. Urtoss Apria, Urtoss Deta, Usamma Spec	radiocnemistry - suite ∠: Gross Apria, Gross Detai, Gamma Spec)																								
		cadiochemistry -	- Suite 2: Gross	Alpha, Gross Beta, Gam	ma Spec																									

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Plan Table Number: LTS_ECM_2005

SAP Number:

Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

The sampling activity displayed on this table represents the first 6 to 9 characters of the sample identification number

DRAFT

Sampler: Haney, T. J.

D - Double OC Volume T - Triple OC Volume

Project Manager: HANEY, T. J./VANHORN, R. L. SMO Contact: MCGRIFF, T. W. Enter Analysis Types (AT) and Quantity Requested Sample Description Sample Location AT1 AT2 AT3 AT4 AT5 AT6 AT7 AT8 AT9 AT10 AT11 AT12 AT13 AT14 AT15 AT16 AT17 AT18 AT19 AT20 Sample Coll Sampling Planned Type of Depth Sampling Sample 3A 9A 1G R5 3Z PH HG 3Y N7 RH RN LA Activity Type Matrix Type Method Location Location (ft) ECT147 REG ANIMAL BIOTA GRAB COMP 06/01/2005 INEEL DEER MOUSE TAN NA TAN 5 GRAB COMP 06/01/2005 DEER MOUSE NA ECT148 REG ANIMAL BIOTA INEEL ECT149 REG ANIMAL BIOTA GRAB COMP 06/01/2005 INEEL DEER MOUSE TAN NA 5 5 ECT150 REG ANIMAL BIOTA GRAB COMP 06/01/2005 INEEL DEER MOUSE TAN NA ECT151 REG ANIMAL BIOTA GRAB COMP 06/01/2005 INEEL DEER MOUSE TAN NA 5 ECT152 REG ANIMAL BIOTA GRAB COMP 06/01/2005 INEEL DEER MOUSE TAN NA ECT153 REG ANIMAL BIOTA GRAB COMP 06/01/2005 INEEL DEER MOUSE TAN NA 5 5 NA ECT154 REG ANIMAL BIOTA GRAB COMP 06/01/2005 INEEL DEER MOUSE TAN NA 5 ECT155 REG ANIMAL BIOTA GRAB COMP 06/01/2005 INEEL DEER MOUSE TAN ECT156 REG ANIMAL BIOTA GRAB COMP 06/01/2005 INEEL DEER MOUSE TAN NA ECT157 REG/QC PLANT BIOTA DUP COMP 06/01/2005 INEEL SAGEBRUSH TAN NA COMP 06/01/2005 SAGEBRUSH ECT158 REG PLANT BIOTA GRAB INEEL TAN NA ECT159 REG GRAB COMP 06/01/2005 SAGEBRUSH TAN NA PLANT BIOTA INEEL ECT160 REG PLANT BIOTA GRAB COMP 06/01/2005 INEEL SAGEBRUSH TAN NA ECT161 REG PLANT BIOTA GRAB COMP 06/01/2005 SAGEBRUSH TAN

AT1: Analysis Suite #1	AT11: Radiochemistry - Suite 2	1000 1	Comments:
AT2: Analysis Suite #2	AT12: Total Metals (TAL)		NODA: Naval Ordnance Disposal Area
AT3: Analysis Suite #3	AT13:		MDA: Mass Detonation Area RAIL: Railcar Explosion area
AT4: Gamma Screen	AT14:		TAN: Test Area North
ATS: Histapthy	AT15:		BLRS: Big Lost River Sinks
All Indupory	ATTO.		B. C.
AT6; Hydrogen Ion (pH)	AT16:		Analysis Suite #3 and gamma screen are field screening tests.
AT7: Mercury	AT17:		Sale Petrovers and the organization of the Control
AT8: Mesoarthopod	AT18:		
AT9: Nitroaromatics (8330)	AT19:		
AT10: Radiochemistry - Suite 1	AT20:		
Analysis Suites:		Contingencies:	
Analysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity			
Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test			
Analysis Suite #3: Mercury, Nitroaromatics (8330)			
Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90	_		
Radiochemistry - Suite 2: Gross Alpha, Gross Beta, Gamma Spec			
Ser insulation with description and a contract and	·		
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The complete sample identification number will appear on the sample labels.

Plan Table Number: LTS_ECM_2005

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SAP Number: Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

Project Manager: HANEY, T. J./VANHORN, R. L.

Sample Sample Sample Sample Sample Sample Type Method Date Type Type		S	Sample Description					Sample I	ocation		L				_	- 1	Enter	Analy:	sis Typ	es (AT) and	Quant	ity Req	quested			_	_
ECT163 REG PLANT BIOTA GRAB COMP 0610005 NEEL SAGEBRUSH TAN NA N			1277 A 1 1 1 7 7 7 1 1 1 1 1 1 1 1 1 1 1 1	(39/4)		200000000000000000000000000000000000000	Area		Location	120 180 180 1	\vdash	⊢			\perp	-	+	+	+	+	+	+	13 AT1	4 AT15	AT16	AT17 A	AT18 A	F194
ECT164 REG PLATE BOTA GRAB COMP 06010005 NEEL SAGEBRUSH TAN NA N	ECT162	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	TAN	NA NA	T	T	Н	H	\forall		1	$^{+}$	t	1	1	t	+		Н	+	+	┪
ECTIEF REG PLANT BOTA GRAB COMP 0610005 NEEL SAGEBRUSH TAN NA N	ECT163	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	TAN	NA NA		Т		П	1		1	T		1:	1		\top		П	T	\top	٦
ECT166 REG PLANT BIOTA GRAB COMP 66110005 NEEL CRESTED WHEATOR TAN NA N	ECT164	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	TAN	NA NA	Г	Т		П	T		1	T	T	1	1			П	П	T	\top	Π
ECTIFF REGIC PLANT BIOTA OUP COMP 661/2005 NEEL CRESTED WHEATOR TAN NA N	ECT165	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	TAN	NA	Г	Г		П	T		1	T	T	1	1				П	7	\top	
ECTIVE REG PLANT BIOTA GRAB COMP 06012005 NEEL CRESTED WHEATOR TAN NA N	ECT166	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	TAN	NA	Г	П	П	П	T		1	T	1	1	1			\Box		T		
ECTIF9 REG PLANT BIOTA GRAB COMP 0601/2005 NEEL CRESTED WHEATOR TAN NA N	ECT167	REG/QC	PLANT BIOTA	DUP	COMP	06/01/2005	INEEL	CRESTED WHEATGR	TAN	NA	Г	П	П	П			2	T	1	2	1		\top	\Box	П	T	\top	_
ECT171 REG PLANT BIOTA GRAB COMP 0601/2005 NEEL CRESTED WHEATGR TAN NA N	ECT168	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	TAN	NA							1	T	1	1	1			П	П	1	\top	
ECT171 REG PLANT BIOTA GRAB COMP 0601/2005 NEEL CRESTED WHEATGR TAN NA N	ECT169	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	TAN	NA	П	П		П	T		1	T	T	1	11		T		П	T	\top	٦
ECT172 REG PLNT BIOTA GRAB COMP 06012005 NEEL CRESTED WHEATOR TAN NA N	ECT170	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	TAN	NA NA	П	П		П	٦		1	T	T	1	- 1							
ECT173 REG PLANT BIOTA GRAB COMP 06/01/2005 INEEL CRESTED WHEATOR TAN NA I I I I I I I I I I I I I I I I I	ECT171	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	TAN	NA	П	П	П	П	T		1	T	T	1	-		T	П	П	T	\top	_
ECT174 REG PLANT BIOTA GRAB COMP 66 01/2005 NEEL CRESTED WHEATGR TAN NA I I I I I I I I I I I I I I I I I	ECT172	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	TAN	NA		П		П	П		1	T	T	1	٠,			П	П	T	\top	
ECT175 REG PLANT BIOTA GRAB COMP 0601/2005 NEEL CRESTED WHEATGR TAN NA 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECT173	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	TAN	NA	П	П		П	П		1	T	T	1	19		\top	\Box	П	T	\top	_
EC176 REG PLANT BIOTA GRAB COMP 0601/2005 INEL CRESTED WHEATGR TAN NA 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECT174	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	TAN	NA NA							1	T		1	1				П	T	\top	
the sampling activity displayed on this table represents the first 6 to 9 characters of the sample identification number. The complete sample identification number will appear on the sample labels. ATI: Radiochemistry - Suite 2 Comments: NODA. Naval Ordnance Disposal Area MDA: Mass Debonation Area MDA: Mass Debonation Area RAIL: Ralion Explosion area TAN: Test Area North BLRS: Big Lost River Sinks Histaphry ATI: Mercury ATI: Analysis Sulte #3 Mercury, Nitroaromatics (6330)	ECT175	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	TAN	NA	П	П		П			1	Т	T	1	3	Π			П			٦
Analysis Sulte #1	ECT176	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	TAN	NA	П	Г		П	T		1	Т	T	1	1		Т	П	П	T	T	Ξ
Mercury	1: Analysis: 2: Analysis: 3: Analysis: 4: Gamma S	Suite #1 Suite #2 Suite #3 Screen	on this table represents th	e first 6 to 9	characters	of the sample ide	ntification number.	AT11: Radiochemistry - Suite 2 AT12: Total Metals (TAL) AT13: AT14:	10077 100	labels.			- !	NODA: MDA: I RAIL: I TAN: T	Nava Mass D Railcar est Ar	Ordnar Detonation Explosi ea North	on Are	sposal a		т.	- Trip	le QC	Volum				<u>=</u>	_
T9; Nitroaromatics (8330) AT19: T10: Radiochemistry - Suite 1 AT20: Inalysis Suites: Contingencies: Inalysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity Inalysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Inalysis Suite #3: Mercury, Nitroaromatics (8330)	T7: Mercury	11-81						AT17:					_	Analysi	s Suite	#3 and	gamm	na scre	en are	field s	creen	ng tes	ts.				\equiv	=
T10: Radiochemistry - Suite 1 Aralysis Suites #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroaromatics (8330)													- :	_									_	_			_	_
Analysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroaromatica (8330)			8										_ :										_				_	_
Analysis Suite #3: Mercury. Nitroaromatics (8330)	malysis Suite#			712000000000000000000000000000000000000	ange Capac	ity		54 9A321131	Contingencies:																			_
Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-lso, U-lso, Sr-90		3: Mercury, Nitro	caromatics (8330)																				_	_	_		_	_
Radiochemistry - Suite 2: Gross Alpha, Gross Beta, Gammu Spec		- Suite 1: Am-24)																							_

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Plan Table Number: LTS_ECM_2005 DRAFT

SAP Number: Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

Project Manager: HANEY, T. J./VANHORN, R. L.

Sempling Sempling		S	Sample Description					Sample t	ocation							1	Enter A	Analysi	з Тур	es (AT)	and C	uantity	Reque	sted		_		_
SCT179 REG SOL GAPA COMP GREL SURFACE SOL TAN Q 2 MCHES 1 Q Q Q Q Q Q Q Q Q	100000000000000000000000000000000000000		100000000000000000000000000000000000000	(2000)			Area	- 28983833	Location	12000000	H	-	_		\perp	-	+	+	+	+		AT13	AT14	AT15	AT16 A	T17 A	T18 AT	19AT2
ECT190 REG SOL GRAB COMP 06010005 NEEL SURFACE SOL TAN 0 2 NOHES 1 0 0 1 0 1 0 0 0 0 0 0 0 0 0 0 0 0 0	ECT177	REG/QC	SOIL	DUP	COMP	06/01/2005	INEEL	SURFACE SOIL	TAN	0-2 INCHES	2	П		П	П	2	Т	Т	2		2	П	T			T	T	Т
ECT191 REG SOL GNAB COMP 06/10006 NEEL SURFACE SOL TAN 0-2 NOHES 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	ECT178	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOIL	TAN	0-2 INCHES	1.					1			1		1	П					T	T
SCT161 REG SOIL GRAB COMP 66912005 NEEL SURFACE SOIL TAN 0.2 NCHES 1 0 0 1 0 1 0 0 0 0	ECT179	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOIL	TAN	0-2 INCHES	1	П			T	1	Т		1		1	П	П		П	T	1	Т
ECT182 REG SOIL GRAB COMP 06012005 NEEL SURFACE SOIL TAN 0.2 NCHES 1 0 0 0 1 0 1 0 1 0 0 0 0 0 0 0 0 0 0	ECT180	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOIL	TAN	0-2 INCHES	1	П		П	П	1	Т		1		1	П	T	П	Т	T	T	T
REG	ECT181	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOIL	TAN	0-2 INCHES	1	П		T	T	1	Т		1		1	П	T	П	T	Ť	Ť	T
ECT146 REG SOL GRAB COMP 661/2005 NEEL SURFACE SOL TAN 0 2 NOIFES 1 0 0 0 1 0 0 1 0 0 1 0 0 0 0 0 0 0 0	ECT182	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOIL	TAN	0-2 INCHES	1	П		\Box	T	1	T	T	1		1	П	T	T	T	†	\dagger	†
ECT195 REG SOIL GRAB COMP 06017005 NEEL SURSURFACE SOIL TAN 0.2 NITES 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	ECT183	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOIL	TAN	0-2 INCHES	1	П		\Box	T	1	T	T	1	1	1	П	T	T	\top	†	Ť	\top
ECT186 REG SOIL GRAB COMP 060172005 NEEL SUBSURFACE SOIL TAN 0 2 NCHES 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECT184	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOIL	TAN	0-2 INCHES	1	П	П	\Box	\forall	1	T	T	1		1	П	T	T	T	1	Ť	\top
ECT187 REGUCC SOIL DU COMP 06017005 NEEL SUBSURFACE SOIL TAN 2-24 NCHES 2 2 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECT185	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOIL	TAN	0-2 INCHES	1	П		\exists	T	1	T	T	1	T	1	П	T		T	1	1	T
ECT168 REG SOIL GRAP COMP 0601/2005 NEEL SUBSURFACE SOIL TAN 2-24 NCHES 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECT186	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOIL	TAN	0-2 INCHES	1	П		П	T	1	T	T	1		1	П	T	T	T	T	†	1
ECT199 REG SOIL GRAB COMP 0601/0005 NEEL SUBSURFACE SOIL TAN 2.24 NCHES 1 I I I I I I I I I I I I I I I I I I	ECT187	REG/QC	SOIL	DUP	COMP	06/01/2005	INEEL	SUBSURFACE SOIL	TAN	2-24 INCHES	2	Н		\dashv	\forall	2	T	T	2	t	2	П	7	7	\top	†	†	\top
ECT190 REG SOIL GRAB COMP 0601/2005 NEEL SUBSURFACE SOIL TAN 2.24 INCHES 1	ECT188	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SUBSURFACE SOIL	TAN	2-24 INCHES	1	П		\dashv	\forall	1	t	T	1		1	П	\forall	\dashv	\top	†	†	†
ECT191 RG SOIL GRAB COMP 0601/2005 NEEL SUBSURFACE SOIL TAN 2-24 NCHES 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECT189	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SUBSURFACE SOIL	TAN	2-24 INCHES	1	Н		\dashv	┪	1	t	T	1	t	1	П	\forall	T	\top	†	†	$^{+}$
The sampling advity displayed on this table represents the first 6 to 9 characters of the sample identification number will appear on the sample labels. AT1: Analysis Suite #1	ECT190	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SUBSURFACE SOIL	TAN	2-24 INCHES	1	Н		\dashv	\forall	1	t	t	1	\vdash	1	H	\forall	\forall	\top	†	†	†
AT1: Analysis Suite #1	ECT191	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SUBSURFACE SOIL	TAN	2-24 INCHES	1	Н		\exists	\forall	1	t	t	1	t	1	H	\forall	T	1	†	†	$^{+}$
	AT1: Analysis 5 AT2: Analysis 5 AT3: Analysis 5 AT4: Gamma S AT5: Histapthy AT6: Hydrogen AT7: Mercury AT8: Mesoarth AT9: Nitroarom AT10: Radioche Analysis Suite #1 Analysis Suite #2 Analysis Suite #2 Analysis Suite #2	Suite #1 Suite #2 Suite #3 Sorreen lon (pH) Spod atics (8330) Moisture Com Earthworm Tc Mercury, Nitre	lent, Hydrogen Ion (pH), xxicity Test, Rye Grass G paromatics (8330)	Cation Exch	ange Capac			AT11: Radiochemistry - Suite 2 Total Metals (TAL) AT13: AT14: AT16: AT17: AT18: AT19:		labels.				NODA: MDA: I RAIL: I TAN: T BLRS:	Mass D Railcar Test Are Big Los	Ordnand etonation Explosion a North at River S	n Area n area inks	oosal A	rea		250		lume					
					0																							

Plan Table Number: LTS_ECM_2005

SAP Number:

Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

DRAFT

Project Manager: HANEY, T. J./VANHORN, R. L.

Sampler: Haney, T. J.

SMO Contact: MCGRIFF, T. W.

Semple S		S	Sample Description					Sample	Location							E	nter A	nalysi	з Туре	es (AT)	and Q	uantity	y Reque	ested				
Activity Type	Т	0.000.0000.000	201 S2	Ι	I	-		Mi Mi		I	AT1	AT2	АТ3	AT4	AT5 A	16 AT	ATE	ATS	AT1	O AT1	AT12	AT13	AT14	AT15	AT16	T17 AT	18 AT	19AT2
ECTISS REG SOL GRAB COMP 06010005 NEEL SUBSUPFACE SOL TAN 2-24 NOVES 1 N 1 N 1 N 1 N 1 N 1 N 1 N 1 N 1 N 1	200.410			(39.5)		A1000000000000000000000000000000000000	Area		Location	100000000000000000000000000000000000000	3A	9A	1G	R5	3Z P	н на	3Y	N7	RH	RN	LA	T	Н	H	7	†	\dagger	t
ECT195 REG SOIL GRAB COUP 06510005 NEEL SUBSURFACE SOIL TAN 224 NCHES 1	192	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SUBSURFACE SOIL	TAN	2-24 INCHES	1	П		П		1	T	Г	1		1			n i		T	T	Т
ECT196 REG SOIL GRAB COMP 06010005 NEEL SUBSURFACE SOIL TAN 2.24 NCHES 1	193	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SUBSURFACE SOIL	TAN	2-24 INCHES	1.	П				1		Г	1		1			J.J.			T	Т
ECT196 REG SOIL GRAB COMP 06112005 NEEL SUBSURFACE SOIL TAN 2-24 NCHES 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	194	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SUBSURFACE SOIL	TAN	2-24 INCHES	1	П		П		1	T	T	1		1	Г	П		T	1	T	T
ECT197 REGOC SOIL DUP COMP 06012005 NEEL SOIL TAN 0.3 NCHES 2 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	195	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SUBSURFACE SOIL	TAN	2-24 INCHES	1	П		П		1	T	Τ	1		1		П			1	T	T
ECT198 REG SOIL GRAB COMP 06/1/2005 NEEL SOIL TAN 0.3 NCHES 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	196	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SUBSURFACE SOIL	TAN	2-24 INCHES	1	П				1	T	T	1	T	1					T	Ť	T
ECT199 REG SOIL GRAB COMP 6601/2005 NEEL SOIL TAN G-3 NCHES G-1 G-	197	REG/QC	SOIL	DUP	COMP	06/01/2005	INEEL	SOIL	TAN	0-3 INCHES	П	П		\Box	\top	T	2	T	T	T	T	П	П		\neg	十	Ť	Ť
ECT200 REG SOIL GRAB COMP 66012005 INEEL SOIL TAN 0-3 INCHES 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	198	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SOIL	TAN	0-3 INCHES	П	П				T	1	T	T	T	T		П	П	\Box	T	Ť	Ť
ECT201 REG SOIL GRAB COMP 0601/2005 NEEL SOIL TAN 0-3 NCHES 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	199	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SOIL	TAN	0-3 INCHES	П	П		\Box	\top	T	1	T	T	T	Г	Г	П	П	\Box	T	T	T
ECT202 REG SOIL GRAB COMP 66/01/2005 INEEL SOIL TAN O.3 INCHES I I I I I I I I I I I I I I I I I I	200	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SOIL	TAN	0-3 INCHES	Г	П		П			1	Τ	T	T	Г	Г				1	T	T
ECT203 REG SOIL GRAB COMP 661/2005 NEEL SOIL TAN 0-3 INCHES 1 1 1 1 1 1 1 1 1	7201	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SOIL	TAN	0-3 INCHES	П	П		П			1	T	Т		Г	Г	П	П	T	T	T	T
ECT204 REG SOIL GRAB COMP 0601/2005 NEEL SOIL TAN 0.3 NCHES 1 1 1 1 1 1 1 1 1	202	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SOIL	TAN	0-3 INCHES		П					1	T	T	1			П			1	T	T
ECT205 REG SOIL GRAB COMP 0601/2005 INEEL SOIL TAN 0-3 INCHES 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	203	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SOIL	TAN	0-3 INCHES	Г	П		П	\forall	\top	1	T	T	T	T	Т	П		\neg	1	Ť	Ť
ECT206 REG SOIL GRAB COMP 06/01/2005 INEEL SOIL TAN 0-3 INCHES 0-1 1	204	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SOIL	TAN	0-3 INCHES	Г	П		П	\forall	T	1	T	T	T	Т	Г	П		T	┰	Ť	T
The sampling activity displayed on this table represents the first 6 to 9 characters of the sample identification number. AT1: Analysis Suite #1 AT2: Analysis Suite #2 AT3: Analysis Suite #3 AT4: Gamma Screen AT4: Total Metals (TAL) AT3: Analysis Suite #3 AT4: Gamma Screen AT5: Histapthy AT16: Bursury AT6: Hydrogen Ion (pH) AT7: Mercury AT8: Mesoarthopod AT18: Analysis Suite #3 AT19: Nitroaromatics (8330) AT19: Analysis Suite #3 AT19: Suite #3 AT19: Marking Suite #3 AT19: Suite #3 AT19: Marking Suite #3 AT19: Marking Suite #3 AT19: Suite #3 AT19: Marking Suite	205	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SOIL	TAN	0-3 INCHES		П		T	\exists	T	1	T	T	T	T	Г	П	\exists		1	†	Ť
AT1: Analysis Sulte #1	206	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SOIL	TAN	0-3 INCHES	Г	П		\exists	1	1	1	T	T	T	Т		П		\neg	\top	T	T
AT12	pling activit	ity displayed o	on this table represents the	ne first 6 to 9	9 characters	of the sample ide	entification number.	The complete sample identification	number will appear on the sample	labels.		_		_		D - Do	uble (C Vo	lume	т-	Triple	QC V	olume				_	_
AT2 Analysis Sulte #2 AT12 Total Metals (TAL) MDA: Mass Detonation Area ATI: Analysis Sulte #3 AT13 RAIL: Railcar Explosion area ATI: Gamma Screen ATI: Total Metals (TAL) MDA: Mass Detonation Area RAIL: Railcar Explosion area TAN: Test Area North BLRS: Big Lost River Sinks BLRS: Big Lost River Sinks AT16 Analysis Suite #3 and gamma screen are field screening tests. AT16 AT16 Analysis Suite #3 and gamma screen are field screening tests. AT18 Mesoarthopod AT18 AT18 AT18 AT18 AT18 AT18 AT19 AT19 AT19 AT19 AT19 AT19 AT19 AT19	Analysis Su	uite #1				(3)		AT11: Radiochemistry - Suite 2	100.00								0200											
AT3. Analysis Suite #3 AT13. Ralicar Explosion area AT4. Garma Screen AT14. Talk. Test Area North AT5. Histapthy AT15. Histapthy AT6. Hydrogen lon (pH) AT16. AT17. Mercury AT7. Mercury AT17. AT18. Mesoarthopod AT8. Mesoarthopod AT18. AT19. Nitroaromatics (8330) AT9. Nitroaromatics (8330) AT19. AT19. AT19. Suites: Contingencies: Contingencies: Analysis Suite #1. Moisture Content, Hydrogen lon (pH), Cation Exchange Capacity Contingencies: Contingencies: Analysis Suite #2. Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3. Mercury, Nitroaromatics (8330) AT19. Analysis Suite #3. Mercury, Nitroaromatics (8330)	Analysis Su	uite #2						AT12: Total Metals (TAL)									_	osal A	rea			_	_					-
AT4: Gamma Screen AT14: TAN: Test Area North AT5: Histapthy AT15: BLRS: Big Lost River Sinks AT6: Hydrogen lon (pH) AT16: Analysis Suite #3 and gamma screen are field screening tests. AT7: Mercury AT17: AT18: AT18: AT8: Mesoarthopod AT18: ————————————————————————————————————	Analysis Su	uite #3						AT13:														_	_					_
AT5: Histapthy AT15: AT15: BLRS: Big Lost River Sinks AT6: Hydrogen lon (pH) AT16: Analysis Suite #3 and gamma screen are field screening tests. AT7: Mercury AT17: Mercury AT18: Searth-pood AT18: Searth-pood AT19: Searth-pood	Gamma Scr	reen						AT14:																				_
AT7: Mercury AT18: Mesoarthopod AT18:	Histapthy							SOLUTION CO.					- 1	BLRS:	Big Lost	River Si	inks					_	_					_
AT7: Mercury AT1:	Hydrogen Id	ion (pH)						AT16:					_ ;	Analysi	s Suite #	3 and g	amma	scree	n are	field sc	reenin	g tests						-
AT19: Nitroaromatics (8330) AT19: AT20: Analysis Suites 1: Contingencies: Analysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroaromatics (8330)	Mercury							AT17:																				_
AT10: Radiochemistry - Suite 1 Analysis Suites: Analysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroaromatics (8330)	Mesoarthop	pod						AT18:						_								_						_
Analysis Suites: Analysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroaromatics (8330)	Nitroaromat	itics (8330)						AT19:					_ :									_						_
Analysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroaromatics (8330)	Radiochemi	nistry - Suite 1	8					AT20:					_ :									_	_					_
Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroaromatics (8330)	Suites:								Contingencies:																			
Analysis Suite #3: Mercury, Nitroaromatics (8330)	Suite #1:1	Moisture Con	itent, Hydrogen Ion (pH),	Cation Exch	ange Capac	city																_						
	s Suite #2: 8	Earthworm To	oxicity Test, Rye Grass G	Browth Test																								
Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90																												_
			CHARLEST AND		0																	_						_
Radiochemistry - Suite 2: Gross Alpha, Gross Beta, Gamma Spec	nemistry - S	Suite 2: Gross	Alpha, Gross Beta, Gam	ma Spec																								_

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Plan Table Number: LTS_ECM_2005

SAP Number: Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

DRAFT

Sampler: Haney, T. J. Project Manager: HANEY, T. J./VANHORN, R. L. SMO Contact: MCGRIFF, T. W.

	S	Sample Description						Sample	Location							Е	nter Ar	nalysis	Types	(AT) a	nd Qua	intity R	equeste	d		
		x6 55	_	_			_	- 246			AT1	AT2	АТЗ	AT4	AT5 AT	6 AT7	AT8	AT9	AT10	AT11	AT12 A	T13 A	T14 AT1	5 AT16	AT17 A	AT18
Sampling Activity	Sample Type	Sample Matrix	Coll Type	Sampling Method	Planned Date	Area		Type of Location	Location	Depth (ft)	3A	9A	1G	R5	3Z Pi	+	\vdash	Н			LA	\dagger	+	T	\dashv	\forall
ECT207	REG/QC	SOIL	DUP	COMP	06/01/2005	INEEL		SOIL	TAN	0-12 INCHES	Г	2		T		T		П		T	T			T	П	寸
ECT208	REG	SOIL	GRAB	COMP	06/01/2005	INEEL		SOIL	TAN	0-12 INCHES		13						П			T					\neg
ECT209	REG	SOIL	GRAB	COMP	06/01/2005	INEEL		SOIL	TAN	0-12 INCHES	Г	1		П			П	П		П	T			Т		\neg
ECT210	REG	SOIL	GRAB	COMP	06/01/2005	INEEL		SOIL	TAN	0-12 INCHES	Г	1	T	T	T	1		П		ヿ	T	1		Τ		ヿ
ECT211	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	1	SOIL	TAN	0-12 INCHES	Г	1		T	\top	T	Г	П	T	\dashv	ヿ	\top	1	T	T	寸
ECT212	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	1	SOIL	TAN	0-12 INCHES	T	1	\exists	\exists		†		П		\dashv	\forall	\top		\top	\dashv	寸
ECT213	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	Ť	SOIL	TAN	0-12 INCHES	T	1	┪	\forall	1	†	П	П	T	\dashv	\dashv	+	1	T	\dashv	寸
ECT214	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	1	SOIL	TAN	0-12 INCHES	T	1	\exists	\forall	\top	†	Т	П	\exists	\dashv	Ť	\top	\top	\top		\dashv
ECT215	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	1	SOIL	TAN	0-12 INCHES	Т	1		\dashv	+	T		П		\exists	\forall	†	1	T	Ħ	\dashv
ECT216	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	+	SOIL	TAN	0-12 INCHES	Н	1	\dashv	\dashv	\top	+		H		\dashv	\dashv	†	+	$^{+}$	7	十
ECT217	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	+	SOIL	IET POND (TAN)	0-6 INCHES	\vdash	Н	┪	\dashv	+	1	Т	Н	7	1	\dashv	+	+	T	7	\dashv
ECT218	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	\dagger	SOIL	IET POND (TAN)	6-24 INCHES	T	H	┪	\dashv	\top	1	Т	Н		1	\forall	\top	+	T	\dashv	寸
ECT219	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	+	SOIL	IET POND (TAN)	24-48 INCHES	T	H	T	\dashv	\top	1		П	T	1	\forall	†	\top	$^{+}$	\dashv	ヿ
ECT220	REG	ANIMAL BIOTA	COMP	GRAB	06/01/2005	INEEL	\dagger	TADPOLE/FROG	TRA COLD WAS PO	NA	T	Н	┪	\forall	\forall	+	Н	Н	1	\dashv	1	+	+	$^{+}$		\dashv
ECT221	REG	ANIMAL BIOTA	COMP	GRAB	06/01/2005	INEEL	+	TADPOLE/FROG	TRA COLD WAS PO	NA NA	H	H	\dashv	\dashv	+	+	Н	Н	1	\dashv	1	+	+	+	\dashv	┪
	tivity displayed o	on this table represents th	e first 6 to 9			ntification number.	_		number will appear on the sample	labels.	_	ш	_			D - Do	ıble Q	C Volu	me	T-T	riple Q	C Volu	me	_		_
1: Analysis					M			Radiochemistry - Suite 2	882				(Comme												
2: Analysis	Suite #2						AT12:	Total Metals (TAL)							Naval O			sal Are	a							
3: Analysis	THE STATE AND LINE						AT13:						-		tailcar E	_										
								-							est Area		area									
4: Gamma							AT14:	<u> </u>							Big Lost F		nks									
5: Histapthy	у						AT15:						- [
6; Hydroge	n Ion (pH)						AT16:						- 2	Analysis	Suite #	3 and g	mma :	screen	are fie	ld scre	ening t	ests.				
7: Mercury							AT17:																			
8: Mesoarti	hopod						AT18:						_ =													
9: Nitroaror	matics (8330)						AT19:	85					ੋ -													
10: Radiochi	emistry - Suite 1	(AT20:						7													
nalysis Suites:								-	Contingencies:																	
San and State of the		tent, Hydrogen Ion (pH),	Cation Exch	ange Capac	itv																					
		oxicity Test, Rye Grass G																								
nalvsis Suite #	3: Mercury, Nitro	paromatics (8330)																								
70000	Suite 1- Am 24	1, Gamma Spec, Pu-Iso,	U-lso, Sr-90)																						
	- Julio 1. MIPZ																									

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Plan Table Number: LTS_ECM_2005

SAP Number: Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

Project Manager: HANEY, T. J./VANHORN, R. L.

	Sample Description					Sample	Location							- 5	Enter	Analy	sis Ty	pes (AT) ar	d Quan	tity Re	quested	ı			
	5/1 55	1							AT1	AT2	AT3	AT4	AT5	AT6 A	17 A	T8 A	T9 A1	T10 A	T11 A	T12 AT	13 AT	14 AT1	5 AT16	AT17	AT18	AT
Sampling Sam Activity Typ		Coll Type	Sampling Method	Planned Date	Area	Type of Location	Location	Depth (ft)	за	9A	1G	R5	3Z	рн н	G 3	Y N	17 F	RH F	RN	A	t	T	\top			r
ECT222 Ri	EG ANIMAL BIOTA	COMP	GRAB	06/01/2005	INEEL	TADPOLE/FROG	TRA COLD WAS PO	NA	Г	Г		П	T		T	T	T	1	1	1			П			Γ
ECT223 RI	EG ANIMAL BIOTA	COMP	GRAB	06/01/2005	INEEL	TADPOLE/FROG	TRA COLD WAS PO	NA		Г						T		1:		1			П	П		Ī
ECT224 RI	EG ANIMAL BIOTA	COMP	GRAB	06/01/2005	INEEL	TADPOLE/FROG	TRA COLD WAS PO	NA	П	Г		П	T	- 0	T	T	1	1	1	1	T	Т	П	П		Γ
ECT225 RI	EG PLANT BIOTA	COMP	GRAB	06/01/2005	INEEL	CATTAIL	TRA COLD WAS PO	NA	Г	Т		T	T		T	T	T	1	T	1	1		П	П		Ī
ECT226 RI	EG PLANT BIOTA	COMP	GRAB	06/01/2005	INEEL	CATTAIL	TRA COLD WAS PO	NA	Т	T		\exists	┪	1	Ť	Ť	†	1	\forall	1	†	\top	Н	П		r
ECT227 RI	EG PLANT BIOTA	COMP	GRAB	06/01/2005	INEEL	CATTAIL	TRA COLD WAS PO	NA NA	Т	T	П	\dashv	\forall	\top	†	†	\dagger	1	\top	1	+	†	Н	П		r
ECT228 RI	EG PLANT BIOTA	COMP	GRAB	06/01/2005	INEEL	CATTAIL	TRA COLD WAS PO	NA		H	П		\forall	+	†	1	+	1	\dagger	1	+	+	Н	Н	_	t
ECT229 RI	EG PLANT BIOTA	COMP	GRAB	06/01/2005	INEEL	CATTAIL	TRA COLD WAS PO	NA	T	t	П	\dashv	\forall	\top	t	†	\dagger	1	\dagger	1		+	Н	П		t
ECT230 RI	EG WATER	COMP	GRAB	06/01/2005	INEEL	PONDWATER	TRA COLD WAS PO	NA NA	Н	\vdash		\dashv	\forall	+	t	t	+	1	+	1	+	+	Н	Н		t
ECT231 RI	EG WATER	COMP	GRAB	06/01/2005	INEEL	PONDWATER	TRA COLD WAS PO	NA NA	H	\vdash	Н	\dashv	\forall	+	$^{+}$	+	+	1	+	1	$^{+}$	+	Н	Н	_	ŀ
40000000 00	EG WATER	COMP	GRAB	06/01/2005	INEEL	PONDWATER	TRA COLD WAS PO	NA NA	H	\vdash	Н	\dashv	+	+	+	+	+	1	+	1	+	+	\vdash	Н	-	ŀ
, Herivatores , As	EG WATER	COMP	GRAB	06/01/2005	INEEL	PONDWATER	TRA COLD WAS PO	NA NA	\vdash	╁	Н	\dashv	+	+	+	+	+	1	+	1	┿	+	Н	Н	_	ŀ
resolutions = e	EG WATER	COMP	GRAB	06/01/2005	INEEL	PONDWATER	TRA COLD WAS PO	NA NA	H	⊢	Н	\dashv	+	+	+	+	+	1	+	1	+	+	Н	Н	_	ŀ
STREET, RE	EG SEDIMENT	COMP	GRAB	06/01/2005	INEEL	SEDIMENT		NA NA	H	╁	H	\dashv	+	1	+	+	+	1	+	-	┿	+	H	Н	_	ŀ
		-			-		TRA COLD WAS PO		H	⊢	Н	4	4	-	+	+	+	4	+	1	+	+	\vdash	Н	_	ŀ
ECT236 RI	EG SEDIMENT	COMP	GRAB	06/01/2005	INEEL	SEDIMENT	TRA COLD WAS PO	NA						1				1		1						
	played on this table represents	the first 6 to 9	characters	of the sample ide		The complete sample identification	557	labels.				Comme		D-D	ouble	QC V	olum/	0	T - Tr	ple QC	Volun	10				
T1: Analysis Suite #1						AT11: Radiochemistry - Suite 2								Ordnan	ce Dis	sposal	Area									
T2: Analysis Suite #2	!					AT12: Total Metals (TAL)								etonatio								-				-
73: Analysis Suite #3	3					AT13:					_]	RAIL: 1	Railcar	Explosio	on are	а										
T4: Gamma Screen						AT14:								a North												_
75: Histapthy						AT15:					_ !	BLRS:	Big Los	t River	Sinks	_	_	_	_			_	—		_	-
T6: Hydrogen Ion (ph	-					AT16:					_ 5	Analysi	s Suite	#3 and	gamm	na scr	een ar	re field	scree	ning te	sts.	_	_			-
T7: Mercury						AT17:					- Table -															
T8: Mesoarthopod						AT18:																_	_			_
T9: Nitroaromatics (8	330)					AT19:					ੋ :											_			_	-
T10: Radiochemistry -	Suite 1					AT20:					7												_			-
Analysis Suites:	APACCO III					CHARLES OF THE CONTROL OF THE CONTRO	Contingencies:															_				-
	ure Content, Hydrogen Ion (pH	Cation Eych	anne Canac	itv																						
	worm Toxicity Test, Rye Grass		ango dapat	7																						-
	ury, Nitroaromatics (8330)																									-
																						_				-
Analysis Suite #3: Mercu	: Am-241, Gamma Spec, Pu-ls	o, U-lso, Sr-9)																							
Analysis Suite #3: Mercu Radiochemistry - Suite 1							19															_	_			

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Plan Table Number: LTS_ECM_2005

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SAP Number: Sampler: Haney, T. J. SMO Contact: MCGRIFF, T. W. Project Manager: HANEY, T. J./VANHORN, R. L. Date: 04/18/2005 Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

	S	ample Description						Sample	Location		L	_	_	_	_	-	Enter A	nalysis	з Туре	es (AT)	and Q	uantity F	leques'	led		_	_
Sampling	Sample	Sample	Coll	Sampling	Planned			Type of		Depth	AT1	AT2	AT3	AT4	AT5	AT6 AT	7 AT8	AT9	AT1	0 AT1	1 AT12	AT13 A	T14 A	T15 AT1	AT17	AT18	AT1
Activity	Туре	Matrix	Type	Method	Date	Area		Location	Location	(ft)	ЗА	9A	1G	R5	3Z	PH HO	3Y	N7	RH	RN	LA						
ECT237	REG	SEDIMENT	COMP	GRAB	06/01/2005	INEEL		SEDIMENT	TRA COLD WAS PO	NA						1			1		1						
ECT238	REG	SEDIMENT	COMP	GRAB	06/01/2005	INEEL		SEDIMENT	TRA COLD WAS PO	NA						1			1		1:						
ECT239	SC+REG (SCREEN + REGULAR SAMPLES)	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	0-2 FT			1	1		1		1	1								
ECT240	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	0-2 FT		П	1	1		T		Π	Π								
ECT241	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	0-2 FT		Г	1	1				П	Π	I	Π						
ECT242	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	0-2 FT	Г	Г	1	1				Г	Γ								
ECT243	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	0-2 FT		Г	1	1				Г	Γ		Π	П					
ECT244	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	0-2 FT	Г	Г	1	1			Т	Г	Т	T	Г	T		T	П		
ECT245	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	0-2 FT	Г	T	1	1				T	T	T				1	П		
ECT246	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	0-2 FT			1	1		100			Γ	Ī		3 1	1	-			
ECT247	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	0-2 FT		Г	1	1	П			Т	Т								
ECT248	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	Ī	SEDIMENT	SINKS	0-2 FT			1	1				Г	T	ı							
ECT249	SC+REG	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	†	SEDIMENT	SINKS	2-4 FT		T	1	1		1		1	1		İ					寸	
. Ø. 8:		n this table represents t	he first 6 to 9	characters	of the sample ide				number will appear on the sample	e labels.		•		Comm	0	D-D	ouble C	C Vol	ume	ं ा-	Triple	QC Vol	ime		'	'	į.
1: Analysis !	C217 - 33							Radiochemistry - Suite 2	2		_					Ordnano	e Disp	osal A	rea								
2: Analysis S	S. C.							Total Metals (TAL)			_				_	etonation											Ξ
3: Analysis S	Suite #3						AT13:									Explosio	n area										_
4: Gamma S	Screen						AT14:	li:								ea North at River S	Caller										_
5: Histapthy							AT15:	H					_ *	DLN3:	big Lo	st ruver 3	EIKS										-
6: Hydrogen	n Ion (pH)						AT16:						_	Analys	is Suite	#3 and	gamma	scree	n are	field sc	reening	tests.					
7: Mercury							AT17:	-					_														_
8: Mesoarth	opod						AT18:						_, ;	_													-
9: Nitroaron	natics (8330)						AT19:						_ :														_
10: Radioche	mistry - Suite 1	(AT20:						_ :														_
nalysis Suites:									Contingencies:																		
nalysis Suite #1	1: Moisture Con	lent, Hydrogen Ion (pH).	Cation Exch	ange Capac	ity																						
		xicity Test, Rye Grass (Growth Test																								_
		paromatics (8330)																									_
		1, Gamma Spec, Pu-Iso)																							_
adiochemistry -	- Suite 2: Gross	Alpha, Gross Beta, Gan	nma Spec																								_
									1000																		_

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Plan Table Number: LTS_ECM_2005

SAP Number: Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

DRAFT

Project Manager: HANEY, T. J./VANHORN, R. L. SMO Contact: MCGRIFF, T. W.

Sampler: Haney, T. J.

	Si	ample Description					Sample	e Location		L					Е	nter Ar	nalysis	Types	(AT) a	nd Qua	ntity R	equeste	d :		
		M M	_	_						AT1	AT2	AT3	AT4	AT5 AT	6 AT7	AT8	AT9	AT10	AT11	AT12 A	T13 A	T14 AT1	5 AT16	AT17 A	T18 AT
Sampling Activity	Sample Type	Sample Matrix	Coll Type	Sampling Method	Planned Date	Area	Type of Location	Location	Depth (ft)	3A	9A	1G	R5	3Z Pi	HG	3Y	N7	RH	RN	LA	\dagger	$^{+}$	H	†	†
	(SCREEN + REGULAR SAMPLES)							2																	
ECT250	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	2-4 FT			1	1		Τ						T		П		
ECT251	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	2-4 FT		П	1:	1:						П	\Box	T	П	П		Т
ECT252	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	2-4 FT		П	1	1		0.00			Г	П	П	Т		П		7
ECT253	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	2-4 FT	Г	П	1	1				Г		П	\exists			П	1	1
ECT254	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	2-4 FT	Г	П	1	1				Г		П	T	T		П	T	T
ECT255	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	2-4 FT	Г	П	1	1		1		Г	Г	П	\exists	\top		П	T	1
ECT256	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	2-4 FT	Г	П	1	1	T	T	Г	Г	Г	П	\Box	\top	T	П	十	\top
ECT257	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	2-4 FT	T	П	1	1	T	T				П	\exists	\top	T	П	T	\top
ECT258	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	2-4 FT	Г	П	1	-1	7			Г		П	\exists	\top		Ħ	T	1
ECT259	SC+REG (SCREEN + REGULAR SAMPLES)	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	4-6 FT			1	1		1		1	1			T				
T1: Analysis : T2: Analysis : T3: Analysis : T4: Gamma :	Suite #1 Suite #2 Suite #3 Screen	n this table represents ti	he first 6 to 9	d characters	of the sample ide		AT11: Radiochemistry - Suite: Total Metals (TAL) AT13: AT14:		e labels.			- :	MDA: I RAIL: I TAN: T		onation xplosion North	Dispo Area area			1-1	riple Q	C Volu	me			
15: Histapthy 16: Hydrogen	Occurren						1866 - 1					_ ;	Analysi	s Suite #	3 and ga	amma	screen	are fie	eld scre	ening t	ests.				
17: Mercury							AT17:													200					
T8: Mesoarth	opod						AT18:					_ ;									_				
	natics (8330)						AT19:					- :									_				
5000	mistry - Suite 1						AT20:						_								_				
unalysis Suites: unalysis Suite #1	1: Moisture Conti	ent, Hydrogen Ion (pH).	Cation Exch	ange Capac	ity			Contingencies:																	
nalysis Suite #2	2: Earthworm To	xicity Test, Rye Grass C																			_				
		aromatics (8330)																							
	Suite 1: Am-24	1, Gamma Spec, Pu-Iso	, U-lso, Sr-90	0																					
		Alpha, Gross Beta, Gam																			_				

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SAP Number:

		ample Description						Samola	Location		l						Enter	Analysi	в Туре	s (AT)	and C	uantity l	Request	ed			
		ampie Description						Saniple	Locatori		AT1	AT2	ΔТ3	AT4	AT5	ATE A	17 A1	ВАТО	AT1	DAT1	1 AT12	AT13	AT14 A	15 AT1	AT17	AT18	AT
Sampling Activity	Sample Type	Sample Matrix	Coll Type	Sampling Method	Planned Date	Area		Type of Location	Location	Depth (ft)	3A		_	Н	3Z	+	+	N7	⊢	RN	+		1				
ECT260	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	4-6 FT	Г	П	1	1	П	8 - 23 m		Т	Г	Г	T		Ϋ́			П	Γ
ECT261	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	4-6 FT			10	1:			, .									П	
ECT262	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	4-6 FT			1	1				Т	Г	Г	Π	П		T		П	Γ
ECT263	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	4-6 FT			1	1					Π	Г		П			П	П	Γ
ECT264	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	T	SEDIMENT	SINKS	4-6 FT		П	1	1	T		T		Т	T	T	П	T		П	П	Γ
ECT265	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	T	SEDIMENT	SINKS	4-6 FT	Г	П	1	1	T			T	T	Τ	T	П		\top	П	\Box	Γ
ECT266	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	T	SEDIMENT	SINKS	4-6 FT			1	1	T			\top	T		T	П				\Box	Γ
ECT267	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	T	SEDIMENT	SINKS	4-6 FT	Г	П	1	1	\exists			T	T	Τ	T	П		T		П	Г
ECT268	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	4-6 FT	Г	П	1	1	T				T	Т	T	П			П	П	Г
ECT269	SC+REG (SCREEN + REGULAR SAMPLES)	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	6-8 FT			1	1		7	1	1	1								
ECT270	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	6-8 FT			1	1					Γ	Γ						П	Г
ECT271	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	6-8 FT	П		1	1				Т	Г	Г	T	П				П	Γ
ECT272	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS		SEDIMENT	SINKS	6-8 FT	Г	П	1	1	T			T	T	T	T			T		\Box	Γ
1: Analysis 2: Analysis	Suite #1 Suite #2 Suite #3 Screen	n this table represents t					AT11: AT12: AT13: AT14: AT15:	Radiochemistry - Suite 2 Total Metals (TAL)	number will appear on the sam				- :	MDA: N RAIL: F	Nava fass (tailcai	l Ordnar Detonation Explosing ea North st River	on Area	posal A				QC Voi					_
6: Hydroge 7: Mercury	n Ion (pH)						AT16: AT17:	-711					_ :	Analysis	Suite	#3 and	gamm	a scree	n are l	field so	reenin	g tests.				_	_
8: Mesoart	hopod matics (8330)						AT18: AT19:						- :													_	_
	emistry - Suite 1	(AT20:						7	-												_	-
nalysis Suites nalysis Suite i nalysis Suite i	1: Moisture Cont 2: Earthworm To	ent, Hydrogen Ion (pH). xicity Test, Rye Grass (aromatics (8330)			ity		03480710		Contingencies:				_														_
	- Suite 1: Am-24	1, Gamma Spec, Pu-Isc	. U-lso, Sr-90	0					2.4.																		

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SAP Number:

Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

Project Manager: HANEY, T. J./VANHORN, R. L.

	s	ample Description					Sample	e Location		L					Er	nter An	alysis 7	Types	(AT) an	d Quant	ity Requ	uested			
		M 8					1		_	AT1	AT2	AT3	AT4	AT5 AT	6 AT7	AT8	AT9	AT10	AT11 A	T12 AT1	13 AT14	AT15	AT16 AT	17 AT	18 AT
sampling activity	Sample Type	Sample Matrix	Coll Type	Sampling Method	Planned Date	Area	Type of Location	Location	Depth (ft)	ЗА	9A	1G	R5	3Z PI	HG HG	3Y	N7	RH	RN I	LA	\dagger		\dagger	\dagger	+
ECT273	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	6-8 FT	Г		1	1	-	0			T	T			1		Т	T
ECT274	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	6-8 FT			10	11					1	\neg						T
ECT275	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	6-8 FT			1	1		7-3				T					Т	T
ECT276	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	6-8 FT	Г		1	1						T					Т	T
ECT277	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	6-8 FT	Г	Г	1	1		T									T	T
ECT278	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	6-8 FT	Г	Г	t	1		T				T					T	\top
ECT279	SC+REG (SCREEN + REGULAR SAMPLES)	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	8-10 FT			1	1		1		1	1							
ECT280	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	8-10 FT	Г		1	1						T				П	T	T
ECT281	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	8-10 FT	Г		1	1									1		\top	\top
ECT282	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	8-10 FT			1	1.		SEC. 2				T	3	ii S	1		T	
ECT283	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	8-10 FT		Г	1	1		1										1
ECT284	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	8-10 FT	Г		1	1	T				T							
ECT285	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	SEDIMENT	SINKS	8-10 FT	T	T	1	1	1	T		T	┪	寸	┪			\top	\top	\top
: Analysis S : Analysis S : Analysis S : Analysis S	uite #1 uite #2	n this table represents th	he first 6 to 9	characters	of the sample ide		The complete sample identification AT11: Radiochemistry - Suite AT12: Total Metals (TAL) AT13:		ole labels.			_ :	MDA: N		onation i	Dispo Area			T - Tri	ple QC	Volume				_
Gamma Se							AT14:							est Area	_	arca									
Histapthy							AT15:					_ !	BLRS: E	lig Lost F	River Sin	ks									_
6: Hydrogen 7: Mercury							AT16:					_	Analysis	Suite #3	3 and ga	mma s	creen a	are fiel	ld scree	ning tes	ts.				<u>=</u>
8: Mesoartho	entre de la constante de						AT18:					- 1													
	atics (8330)						AT19:																		
	nistry - Suite 1						AT20:	A Proposition					_					_							_
alysis Suite #2 alysis Suite #3 diochemistry -	Earthworm To Mercury, Nitro Suite 1: Am-24	ent, Hydrogen Ion (pH), exicity Test, Rye Grass G earomatics (8330) 1, Gamma Spec, Pu-Iso Alpha, Gross Beta, Gam	Browth Test		ity			Contingencies:																	<u>=</u>

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Plan Table Number: LTS_ECM_2005

SAP Number: Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

Project Manager: HANEY, T. J./VANHORN, R. L.

Service Serv		S	ample Description						Sample	Location		L	_	_	_	_	Е	nter Ar	alysis	Туре	(AT)	and Qu	uantity	Reque	sted	_	_	_
Control Cont				(250)		*************		Τ	- 15753533	V	100 March 100 Ma						+	-					AT13	AT14	AT15 A	T16 AT	17 AT1	18 AT1
ECTIZE SCREEN SEEMENT COMP GRAB 06110000 BLRS SEEMENT SANSS B-19 TT 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1		3,371,0	(0.0000000)			53,550.0	100000	-	D.SECONORIO	(See 2000, 2001)	1000	3A	9A	-	\vdash	3Z Pi	HG	3Y	N7	RH	RN	LA		Н	+	+	+	+
ECTION SCREEN SERVINET COMP GRAB GOSTOOM BLRS SEDMENT SINKS B-HOFT N N N N N N N N N N N N N N N N N N N			W. C. SW	3555	100			+		3200	52555	H		- 67	-	_	+	-		L	Н	_		\dashv	+	+	+	+
ECX12 REG ANNAL BOTA 0RAB COMP 06010005 NEEL DEERMOUSE MOA NA	ECT287	SCREEN	SEDIMENT		GRAB	06/01/2005	BLRS	+	SEDIMENT	SINKS	8-10 FT	L	L	1	1	4	-	┡	Ш	L	Ш	_	Ц	_	4	+	+	+
ECX12 REG ANIMAL BOTA GRAB COMP 06010005 NREL DEFRMOUSE MCA NA	ECT288	SCREEN	SEDIMENT	COMP	GRAB	06/01/2005	BLRS	ļ_	SEDIMENT	SINKS	8-10 FT	L	Ш	1	1	4	\perp	\perp	Ш	L	Ш	Ц	Ц	\perp	4	4	\perp	丰
ECX123 REG ANIMAL BOTA GRAB COMP 061/2005 NEEL DEER MOUSE MDA NA	ECX121	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL		DEER MOUSE	MDA	NA NA					5	1	Ш	1	1		1						\perp
ECX124 REG	ECX122	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL		DEER MOUSE	MDA	NA					5	1		1	1		1						
ECX125 REG ANIMAL BIOTA GRAB COMP 06012005 NEEL DEER MOUSE MDA NA I I I I I I I I I I I I I I I I I I	ECX123	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL		DEER MOUSE	MDA	NA NA					5	1		1	1		1					Т	Τ
ECX12 REG ANIMAL BIOTA GRAB COMP 06912005 NEEL DEER MOUSE MDA NA I I I I I I I I I I I I I I I I I I	ECX124	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL		DEER MOUSE	MDA	NA NA	П			П	5	1	П	1	1		1					T	T
ECX127 REG ANIMAL BIOTA GRAB COMP 0601/2005 NEEL DEER MOUSE MDA NA I S S I I S I S I I S I I S I S I I S I	ECX125	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	T	DEER MOUSE	MDA	NA NA	П	П		T	5	1	П	1	1	П	1					Т	T
ECX128 REG ANIMAL BIOTA GRAB COMP 0601/2005 NEEL DEER MOUSE MDA NA I 5 5 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECX126	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	1	DEER MOUSE	MDA	NA NA				П	5	1		1	1	П	1			T		T	T
ECX129 REG ANIMAL BIOTA GRAB COMP 0601/2005 INEEL DEER MOUSE MDA NA I 5 5 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECX127	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	T	DEER MOUSE	MDA	NA NA	П	Г	П	П	5	1	П	1	1	П	1	П		T	T	T	\top
ECX131 REGICC PLANT BIOTA GRAB COMP 0601/2005 INEEL SAGEBRUSH NODA NA 5 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECX128	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	1	DEER MOUSE	MDA	NA NA		Г		П	5	1	Г	1	1	П	1			1	\top	1	\top
ECX131 REGICC PLANT BIOTA DUP COMP 66 01/2005 NEEL SAGEBRUSH NODA NA 1 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2	ECX129	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	\top	DEER MOUSE	MDA	NA NA	П	Г	П	T	5	1	Т	1	1	П	1	П	\neg	\top	\top	\top	\top
ECX132 REG PLANT BIOTA GRAB COMP 06/01/2005 INEL SAGEBRUSH NODA NA 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECX130	REG	ANIMAL BIOTA	GRAB	COMP	06/01/2005	INEEL	1	DEER MOUSE	MDA	NA NA	Г	Г	П	T	5	1	П	1	1	П	1	П	\neg	T	\top	\top	\top
es amplier advivity displayed on this table represents the first 6 to 9 characters of the sample identification number will appear on the sample labels. D - Double QC Volume T - Triple QC Volume T - Triple QC Volume AT11: Radiochemistry - Suite 2 Comments: NOA: Navail Ordnance Disposal Area MOA: Mass Detonation Disposal Area MOA: Mass Detonation Disposal Area AT12: Total Metals (TAL) MOA: Mass Detonation Disposal Area MOA: Mass Detonation Disposal Area AT14: Total Metals (TAL) TAN: Test Area North BLRS: Big Lost River Sinks TAN: Test Area North BLRS: Big Lost River Sinks Analysis Suite #3 and gamma screen are field screening tests. AT17: Mercury Mercur	ECX131	REG/QC	PLANT BIOTA	DUP	COMP	06/01/2005	INEEL	1	SAGEBRUSH	NODA	NA NA	П	Г	П	T	T	2	Т	2	2	П	2	П	\neg	\top	T	†	\top
Aralysis Suite #1	ECX132	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	1	SAGEBRUSH	NODA	NA NA	Г	Г		T		1	T	1	1	П	1	П	\neg		T	T	†
AT14: TAN: Test Area North BLRS: Big Lost River Sinks Histapthy AT15: Analysis Suite #3 and gamma screen are field screening tests. Analysis Suite #3 and gamma screen are field screening tests. Analysis Suite #3 and gamma screen are field screening tests. Analysis Suite #3 and gamma screen are field screening tests. Analysis Suite #3 and gamma screen are field screening tests. Confingencies: Confingencies: AT20: Confingencies: AT30: Confingencies: AT41: Confingencies: Analysis Suite #1: Moisture Content, Hydrogen lon (pH), Cation Exchange Capacity Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Analysis Suite #3: Mercury, Nitroaromatics (8330) adiochemistry - Suite 1: Am-241, Gamma Spec, Pu-lso, U-lso, Sr-90	1: Analysis 2: Analysis	Suite #1 Suite #2	on this table represents th	e first 6 to 9	characters	of the sample ide	ntification number.	AT11: AT12:	Radiochemistry - Suite 2	1000	ple labels.			_ ;	NODA:	Naval O Mass Det	rdnance	Dispo Area			т-1	riple	QC V	lume				
Histapthy AT15: Histapthy AT16: Analysis Suite #3 and gamma screen are field screening tests. Histapthy Analysis Suite #3 and gamma screen are field screening tests. Analysis Suite #3 and gamma screen are field screening tests. Analysis Suite #3 and gamma screen are field screening tests. Analysis Suite #3 and gamma screen are field screening tests. Confingencies: Confingencies: AT20: Confingencies: AT30: Confingencies: AT30: Confingencies: AT30: Confingencies: AT30: Confingencies: AT30: AT30: Confingencies: AT30: AT30: Confingencies: AT30:								-					_				area										_	
Hydrogen lon (pH) AT16: Analysis Suite #3 and gamma screen are field screening tests. AT17: Mercury Mesoarthopod AT18: Signaturhopod AT19: Nitroaromatics (8330) AT19: Contingencies: AT20: Contingencies: AT20: AT20: AT30:	-								ш.									nks										=
Mesoarthopod	9 7000	/Superson							H					-	Δnalvsi	e Suite #	S and a	amma	creen	are 6	eld sco	enine	toote					_
99: Nitroaromatics (8330) AT19: 110: Radiochemistry - Suite 1 AT20: AT20: Contingencies: AT39: Contingencies: AT39: AT49: AT40: T40: AT40: AT40: AT							- 10		i a					T :	uninya	o Guille in	o an or gr		90,000	are n	cra sor	J. Carrier Land	tosta					
10: Radiochemistry - Suite 1 A720: Contingencies: Inalysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity Inalysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test Inalysis Suite #3: Mercury, Nitroaronatics (8330) adiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90	8: Mesoarth	opod						AT18:							_													_
nalyais Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity nalyais Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test nalyais Suite #3: Mercury, Nitroaromatics (8330) adiochemistry - Suite 1: Am-241, Gamma Spec, Pu-lao, U-lao, Sr-90	9: Nitroaron	natics (8330)						AT19:						_ :														_
nalysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity nalysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test nalysis Suite #3: Mercury, Nitroaromatics (8330) addiochemistry - Suite 1: Am-241, Gamma Spec, Pu-lso, U-lso, Sr-90	10: Radioche	mistry - Suite 1						AT20:	· ·						_													
nalysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test nalysis Suite #3: Mercury, Nitroaromatics (8330) adiochemistry - Suite 1: Am-241, Gamma Spec, Pu-lso, U-lso, Sr-90	Samuel Astronomy									Contingencies:																		
nalysis Suite #3: Mercury, Nitroaromatics (8330) adiochemistry - Suite 1: Am-241, Gamma Spec, Pu-lao, U-lao, Sr-90					ange Capac	rity																						_
adiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90		the state of the s		rowth Test																								_
				U-lso Sr-90	j .					_																		_
adiochemistry - Suite 2: Gross Alpha, Gross Beta, Gamma Spec																												_

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Plan Table Number: LTS_ECM_2005

SAP Number: Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

Project Manager: HANEY, T. J./VANHORN, R. L.

	s	ample Description					Sample Lo	ocation		L				_	En	nter An	alysis	Types	(AT) a	nd Qu	entity R	equest	ed .		_	_
Sampling Activity	Sample Type	Sample Matrix	Coll Type	Sampling Method	Planned Date	Area	Type of Location	Location	Depth (ft)	AT1		AT3	AT4	AT5 AT	+					AT12	T13 A	T14 AT	15 AT16	AT17	T18 A1	19
ECX133	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	NODA	NA	T	П	П	\exists		1	П	1	1	\exists	1	1	1		П	\top	7
ECX134	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	NODA	NA		П				1		1	1	\Box	1					T	7
ECX135	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	NODA	NA	Г	П		П		1		1	1	П	1				П	T	٦
ECX136	REG/QC	PLANT BIOTA	DUP	COMP	06/01/2005	INEEL	CRESTED WHEATGR	NODA	NA	Г	П		T		2		2	2	T	2				П	T	
ECX137	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	NODA	NA	Т	П		\exists		1		1	1	\exists	1					\neg	7
ECX138	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	NODA	NA	T	П	П	\exists	T	1	П	1	1	寸	1	\top	T	Т	П	\top	7
ECX139	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	NODA	NA	Т	П		\Box		1		1	1	\Box	1	\top	T		\Box	\top	7
ECX140	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	NODA	NA	T	П	П	\exists	T	1		1	1	\exists	1	\top	T		П	\top	7
ECX141	REG/QC	SOIL	DUP	COMP	06/01/2005	INEEL	SURFACE SOIL	NODA	0-2 INCHES	2	П	П	\exists	T	2		2	2	T	2				П	\top	1
ECX142	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOIL	NODA	0-2 INCHES	1	П	П	\dashv	1	1	П	1	1	\dashv	1	\top	1	T	П	\top	=
ECX143	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOIL	NODA	0-2 INCHES	1	Н	П	\dashv	+	1	Г	1	1	\dashv	1	+	\dagger	1	\vdash	\top	4
ECX144	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOIL	NODA	0-2 INCHES	1	Н	П	ヿ	\top	1	П	1	1	\dashv	1	\top	\top	T	H	\top	_
ECX145	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SURFACE SOIL	NODA	0-2 INCHES	1	H	П	\dashv		1		1	1	\dashv	1	\top	\top	T	\dashv	\top	-
ECX146	REG/QC	SOIL	DUP	COMP	06/01/2005	INEEL	SUBSURFACE SOIL	NODA	2-24 INCHES	2	H	П	\forall	\top	2		2	2	┪	2	\top	\top	$^{+}$	H	\top	7
ECX147	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SUBSURFACE SOIL	NODA	2-24 INCHES	1	Н	H	\dashv		1	Н	1	1	\dashv	1	+	+		\forall	+	Ħ
he sampling activity T1: Analysis Sui T2: Analysis Sui T3: Analysis Sui T4: Gamma Scri T5: Histapthy T6: Hydrogen lo T7: Mercury	ite #1 ite #2 ite #3	n this table represents th	e first 6 to 9	characters	of the sample ide		The complete sample identification in AT11: Radiochemistry - Suite 2 Total Metals (TAL) AT13: AT14: AT15: AT16: AT17:	umber will appear on the sample	a labels.			- <u>!</u>	MDA: M RAIL: R FAN: To BLRS: E		plosion A North Liver Sin	Dispo Area area ks	sal Are	sa .	T - Ti			ime				_ _ _
Mesoarthopo	ics (8330)						AT18:					- :														_
	Moisture Cont	lent, Hydrogen Ion (pH),	711000000000000000000000000000000000000	ange Capac	ity		AT20:	Contingencies:				_														_
	A COMPANY OF THE REAL PROPERTY.	xicity Test, Rye Grass G paromatics (8330)	rowth Test																							-
			II lee Ce Of																							_
Radiochemistry - Su	uite 1: Am-24	I, Garrina Spec, Fu-iso,	0-180, 31-30															_						_	_	

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SAP Number: Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

Project Manager: HANEY, T. J./VANHORN, R. L.

Date: 04/18	2003	Plan Table Revision:			ING TERM ECO				Project Manager: HANE																		
	S	ample Description						Sample	Location							. 1	Enter /	Analysi	з Туре	s (AT)	and Qua	antity Re	queste	j.			
		20 32		_			_	86			AT1	AT2	АТЗ	AT4	AT5	AT6 AT	7 AT	8 AT9	AT10	AT11	AT12	T13 AT	14 AT1	5 AT16	AT17 A	T18 A	119
Sampling Activity	Sample Type	Sample Matrix	Coll Type	Sampling Method	Planned Date	Area		Type of Location	Location	Depth (ft)	3A	9A	1G	R5	3Z	рн но	+	+	⊢		LA	$^{+}$	+	H	\dagger	\dagger	-
ECX148	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	1	SUBSURFACE SOIL	NODA	2-24 INCHES	1	П		T	T			1	1		1		11	П	7	T	Т
ECX149	REG	SOIL	GRAB	COMP	06/01/2005	INEEL		SUBSURFACE SOIL	NODA	2-24 INCHES	1.	П			\top	1		1	1		1:			П		\top	_
ECX150	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	1	SUBSURFACE SOIL	NODA	2-24 INCHES	1	П		T	T	1		1	1	П	1			П	7	\dashv	Π
ECX151	REG/QC	SOIL	DUP	COMP	06/01/2005	INEEL	1	SOIL	NODA	0-3 INCHES	Г	П		T	T		2	T	Г	П	П			П	\neg	\neg	
ECX152	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	1	SOIL	NODA	0-3 INCHES	Г	П		\dashv	\forall	\top	1	T	T	T	Ħ	\top	T	П	十	\top	_
ECX153	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	1	SOIL	NODA	0-3 INCHES	Г	П	П	ヿ	\forall	\top	1		T	Т	\Box	\top	T	П	寸	\top	Τ
ECX154	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	Ť	SOIL	NODA	0-3 INCHES	П	П		\Box	\forall	1	1	t	T		\Box	1		П	\top	\top	٦
ECX155	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	\top	SOIL	NODA	0-3 INCHES	Г	П	П	\forall	\forall	\top	1	T	T	П	\Box	\top	T	П	\top	\top	7
ECX156	REG/QC	SOIL	DUP	COMP	06/01/2005	INEEL	1	SOIL	NODA	0-12 INCHES	Г	2		\exists	7	1	T	T	Т	П	П	1	T	П	1	\top	
ECX157	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	†	SOIL	NODA	0-12 INCHES	Г	1		\dashv	\forall	+	T	T	T	Т	П	1	1	П	十	\dagger	=
ECX158	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	1	SOIL	NODA	0-12 INCHES	Г	1		\dashv	\forall	1	T	T	T	T	\exists	\top	T	П	7	\top	_
ECX159	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	\top	SOIL	NODA	0-12 INCHES	Г	1		ヿ	\forall	\top	Ť	T	T	T	\Box	\top	T	H	寸	\top	_
ECX160	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	1	SOIL	NODA	0-12 INCHES	Г	1		寸	T	\top	T	T	T		Ħ	\top	T	П	十	\top	ī
ECX161	REG/QC	PLANT BIOTA	DUP	COMP	06/01/2005	INEEL	1	SAGEBRUSH	RAIL	NA NA	2	П	П	寸	ヿ	1	2	2	2	Т	2	\top	T	П	\dashv	\top	7
ECX162	REG	PLANT BIOTA	GRAB	COMP	06/01/2005	INEEL		SAGEBRUSH	RAIL	NA NA	1	П		T	\forall	١,		1	1	П	1		1	П	\neg	\top	_
e sampling ac	tivity displayed o	n this table represents th	e first 6 to 9	characters	of the sample ide	entification number.	The com	nplete sample identification	number will appear on the sample	labels.	_	_				D-D	ouble	QC Vo	ume	т-	Triple C	C Volur	ne				_
1: Analysis	Suite #1				(8)		AT11:	Radiochemistry - Suite 2	0.600					Comme													
2: Analysis	Suite #2						AT12	Total Metals (TAL)						_		Ordnani			rea								_
3: Analysis	Suite #3						AT13:						_]	RAIL: R	ailcar	Explosio	n area										
4: Gamma	Screen						AT14:									a North											_
5: Histapthy	y						AT15:						_ !	BLRS: E	ig Los	t River S	Sinks										-
6; Hydroge	n lon (pH)						AT16:	40)					_ 7	Analysis	Suite	#3 and	gamma	a scree	n are f	ield scr	eening	ests.					_
7: Mercury							AT17:																				Ξ
8: Mesoarti	hopod						AT18:	,					-	_													_
9: Nitroaror	matics (8330)						AT19:	8					ੋਂ •														-
	emistry - Suite 1	(AT20:	-					7														_
nalysis Suites:	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,								Contingencies:																		Τ
nalysis Suite #	1: Moisture Con	lent, Hydrogen Ion (pH),	Cation Exch	ange Capaci	ity																						
	CONTRACTOR OF STREET	xicity Test, Rye Grass G	rowth Test																								_
		paromatics (8330)																									_
	 Suite 1: Am-24 	1, Gamma Spec, Pu-Iso,)																							_
	0.4.0.0	Alpha, Gross Beta, Gami	C-																								

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Project Manager: HANEY, T. J./VANHORN, R. L.

Analysis Suite #2 Analysis Suite #3 Anal		s	ample Description					Sample L	ocation		L			_		En	ter An	alysis	Types	(AT) a	nd Qu	antity R	lequest	ed .		_
Second Reco Soil Gree Cours General Cours Second Cou	554 (A.M. 164 W. 16			(39.5)		400000000000000000000000000000000000000				120 100 100 00	\vdash			Н		\Box					\dashv	AT13 A	T14 AT	15 AT16	AT17	AT18 AT
ECTION REG SOL GRAB COMP 06110000 REEL SAMEBRUSH RAA NA 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1			(1000000000			535,500	100(35)	D. SERVICE V	()************************************	1000	\vdash	9A	1G	R5	3Z PH	HG	3Y	\vdash		RN	LA	4	+		\dashv	+
ECK166 REG 0 50L 0RM 0010005 NEEL 0KESTED MEATOR PAAL NA 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	200000	1997	25.00	332			N. Sales	5195 (5199) 20	2000	2.8%	-	Н		Н	+	1		ं	-	Н	1	4	+	+	4	+
ECX166 REGOC SOIL OUP COMP 06010005 NEEL CRESTED WHEATOR RAM NA 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECX164	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	RAIL	NA NA	1.	Ш	\perp	Ц		1		1.	1:	Ш	1	4	4	\perp	_	\bot
ECNIFY REG SOIL GRAB COMP 0617005 NEEL CRESTED WHEATOR RAL NA 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECX165	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SAGEBRUSH	RAIL	NA	1					1		1	1		1					
ECX168 REG SOL GRAB COMP 06112005 NEEL CRESTED WHEATOR RAL NA 1 N N 1 N N N N N N N N N N N N N N	ECX166	REG/QC	SOIL	DUP	COMP	06/01/2005	INEEL	CRESTED WHEATGR	RAIL	NA	1					1		1	1		1					
ECX170 REG SOIL GRAB COMP 06012005 NEEL CRESTED WHEATOR RALL NA 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	ECX167	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	RAIL	NA	1			П		1		1	1		1	T				Т
ECX170 REG SOIL GRAB COMP 0661/2005 NEEL SURFACE SOIL RAIL O.2 NOHES 2 0 0 1 0 1 0 1 0 1 0 0 0 0 0 0 0 0 0 0	ECX168	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	CRESTED WHEATGR	RAIL	NA	1	П	П	П		1		1	1		1	\Box				\top
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SAP Number:

Date: 04/18/2005

Plan Table Revision: 0.0 Project: LONG TERM ECOLOGICAL MONITORING

Project Manager: HANEY, T. J./VANHORN, R. L.

	S	ample Description					Sample	Location		L		_	_	_	Er	nter An	nalysis	Types	(AT) a	nd Qu	antity R	equest	ed			_
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ECX182	REG	SOIL	GRAB	COMP	06/01/2005	INEEL	SOIL	RAIL	0-3 INCHES	Г	П		\exists		T	1		П	T	T			T	П	\neg	_
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Date: 04/18/2005

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	S	ample Description					Sample	Location							E	nter An	alysis '	Types (AT) and	d Quant	ity Req	uested	E			
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Activity	Type	Matrix	Type	Method	Date	Area	Location	Location	(ft)	ЗА	9A	1G	R5 3	Z PH	HG	3Y	N7	RH I	RN L	A			ш			
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The sampling activity displayed on this table represents the first 6 to 9 characters of the sample identification number.	The complete sample is	entification number will appear on the sample labels.	D - Double QC Volume T - Triple QC Volume
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AT2: Analysis Suite #2	AT12: Total Metals	TAL)	NODA: Naval Ordnance Disposal Area
AT3: Analysis Suite #3	AT13:		MDA: Mass Detonation Area RAIL: Railcar Explosion area
AT4: Gamma Screen	AT14:		TAN: Test Area North
AT5: Histapthy	AT15:		BLRS: Big Lost River Sinks
And the state of t	AT16:		
AT6: Hydrogen Ion (pH)	***		Analysis Suite #3 and gamma screen are field screening tests.
AT7: Mercury	AT17:		
AT8: Mesoarthopod	AT18:		"
AT9: Nitroaromatics (8330)	AT19:		
AT10: Radiochemistry - Suite 1	AT20:		
Analysis Suites:		Contingencies:	
Analysis Suite #1: Moisture Content, Hydrogen Ion (pH), Cation Exchange Capacity			
Analysis Suite #2: Earthworm Toxicity Test, Rye Grass Growth Test			
Analysis Suite #3: Mercury, Nitroaromatics (8330)			
Radiochemistry - Suite 1: Am-241, Gamma Spec, Pu-Iso, U-Iso, Sr-90			
Radiochemistry - Suite 2: Gross Alpha, Gross Beta, Gamma Spec		(1) ·	
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Appendix B Sample Collection Procedures

Appendix B

Sample Collection Procedures

B-1. OVERVIEW

Sampling for long-term ecological monitoring (LTEM) occurs as presented in the *Long-Term Ecological Monitoring Plan for the Idaho National Engineering and Environmental Laboratory* (INEEL 2004). Efforts are directed at sampling to determine levels of contamination in the selected media and to detect possible effects. Levels of contamination in soil, deer mice, and plants are determined to validate the Operable Unit (OU) 10-04 ecological risk assessment's assumption of no migration of contamination off the areas of concern (AOCs) and to establish a baseline. Effects data are evaluated for soil fauna, plants, mammals, and avian receptors at the AOCs. This appendix presents the sampling procedures used to collect analytical and effects samples at each AOC:

- 1. Randomly select plots (generally 10) in the site location grids designated for Fiscal Year (FY) 2005 sampling.
- 2. Prepare the plots by staking the corners and center and distributing mammal traps in 3-m (10-ft) intervals on the 100 × 100-m (110 × 110-yd) plot, as shown in Figure B-1 and discussed in Technical Procedure (TPR) -145, "Biotic and Proximal Soil Sampling."

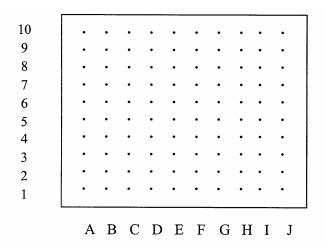


Figure B-1. Example of the transect design.

- 3. Obtain necessary paperwork, including safe work permits, scientific/trapping collection permits, and radiological work permits.
- 4. Obtain all sampling equipment, forms, and labels (as required).
- 5. Sample from May to September 2005:
 - a. Perform soil sampling for plant and earthworm bioassays, analytical concentrations, and soil fauna community structure determination with the Berlese funnel extraction procedure.

- b. Collect plant tissue for analysis.
- c. Sample the small mammal community structure, presence/absence, diversity/richness, and density/biomass using the trap and release methodology (the sampling procedure is presented in Section B-3.1.3).
- d. Sample the plant community structure, presence/absence, diversity/richness, and density/biomass (the sampling procedure is presented in Section B-3.1.1).
- e. Sample bird community structure, presence/absence, diversity/richness, and density/biomass (the sampling procedure is presented in Section B-3.1.2).
- f. Sample deer mouse tissue to obtain effects and analytical data (mice should be collected on the last day of community sampling).
- g. Harvest small mammals for analytical concentration determination (the sampling procedure is presented in TPR-145).
- h. Harvest small mammals for organ-to-body weight measurements, histopathology, and genetic samples (the sampling procedure is presented in Section B-3.4).
- 6. Decontaminate sampling equipment, the task site, and personnel (as necessary).
- 7. Prepare samples for storage and shipment to the appropriate facilities:
 - a. Histopathology specimens will be shipped to the laboratory.
 - b. Preserved invertebrates will be sent to the laboratory.
 - c. Bioassay soils will be shipped to the laboratory for plant and earthworm toxicity bioassays.
 - d. Soil samples will be shipped to the laboratory for chemical and radiological analysis.
 - e. Plant and small mammal samples will be frozen and shipped to the laboratory for chemical and radiological analysis.
 - f. Soil fauna will be extracted and the extract will be shipped to the analyists.

B-2. ANALYTICAL SAMPLING PROCEDURES

B-2.1 Biota Analytical Samples

Samples of vegetation, mammals, and soil will be collected for analysis of contaminant concentration.

B-2.1.1 Vegetation Sampling Procedure for Analytical Sampling

Two types of vegetation (shrubs and grasses), representing the two most common functional plant types at the Idaho National Laboratory (INL), will be collected for chemical analysis. A review of dietary information for herbivorous and omnivorous INL wildlife species has resulted in consideration of the following individual plant species and/or types:

- Wyoming big sagebrush (*Artemisia tridentata*)
- Crested wheatgrass (Agropyron cristatum) or Indian rice grass (Oryzopsis hymendoides)
- Hardstem bulrush (*Scirpus acutus*) or other aquatic plant.

Sagebrush represents the shrub most commonly used by the INL's primary consumers, including the pronghorn, sage grouse, black-tailed jackrabbit, Nuttall's cottontail rabbit, and the pygmy rabbit. In addition, sagebrush is an important component in the diets of avian and mammalian omnivores and herbivorous insects. Wheatgrasses are most widely used and are significant components in the diets of jackrabbits, cottontail rabbits, birds, and small mammals. If crested wheatgrass is unavailable, or the amount is not sufficient, Indian rice grass or other wheatgrasses will be substituted. Hardstem bulrush nutlets are an important waterfowl and shorebird food, while muskrats and geese eat the rhizomes and stems.

Terrestrial vegetation samples will be collected during the early part of the growing season in conjunction with small mammal population analysis and tissue collection. Grass and sagebrush will be sampled in late May or June.

A field reconnaissance will be used to assess species presence and abundance within each randomly selected 100×100 -m (110×110 -yd) grid. If wheatgrass or sagebrush is unavailable, the nearest grid that contains a sufficient amount of these species will be evaluated. A field reconnaissance of potential reference areas also will be completed to match the reference area with the site areas to the greatest extent possible. Final selection of the reference area and sampling grid cells will be based on the presence of suitable species and access.

Each vegetation tissue sample will be a composite of material from at least five individual plants of the same species. Individual plants should be randomly selected from within each 100×100 -m (110×110 -yd) grid. Plants sampled should be distributed across the plot if possible. Atypical individuals (i.e., resembles less than 5% of the plants for the area) based on size or herbivory should not be included. If possible, approximately equal amount of vegetation should be collected from each individual plant.

Clean disposable gloves should be worn. Plant samples should be clipped with pruning shears or grass shears (as appropriate). Plant material from each of the five radial plots should be combined into one plastic bag to make a composite sample. Sagebrush should be clipped on at least two sides and at two different heights to obtain a representative sample.

A minimum weight of fresh biomass required for each analysis is to be provided in the Appendix A field guidance forms. If munitions analyses or other analysis are required, additional sampling may be required. Sample weight should be verified in the field to ensure that an adequate quantity has been collected. Plant samples should be placed into a sealable plastic bag that has been placed into another sealable plastic bag. Sharp points on woody vegetation should be bent or broken off within the bag to avoid bag puncture. Bags should be labeled, and the field data should be recorded in notebooks or on field data sheets. Samples should be placed in a cooler on ice until frozen or shipped to the laboratory. Field data will be recorded.

Grass samples should be collected by clipping above ground level (e.g., 1.3 to 5.1 cm [e.g., 0.5 to 2 in.]) with grass shears. Clipping should be adjusted, as needed, to minimize sampling dead vegetation from previous years and to maximize sampling green vegetation from the current growing season. All material above the cutting height will be collected. Dead material should be removed from the sample by hand if unavoidably collected. Grass samples will include new growth of leaves, stems, and any

inflorescences present on the plants. It is desirable to remove as much dead material as possible; however, this might be impractical, and an estimate of the percentage of dead material should be noted.

Shrub samples should be collected using pruning shears. Collected material will include leaf and stem growth from the current season. Shrubs should be clipped at a height between 0.5 and 1.5 m (0.55 and 1.6 yd) on at least two sides. It is common to also collect woody material during this process. Stripping fresh leaves and stems from the woody material might be necessary. In the event that woody material is not removed, the sampler should make an estimate of the remaining amount.

Macrophytic aquatic plants should be collected along the margins of the wastewater ponds and the Big Lost River sinks. One composite sample will be collected at each aquatic sample location. The aboveground portion of each plant should be cut and placed in a labeled, heavy-duty plastic bag and then placed in a cooler with ice for transport to the analytical laboratory.

These procedures can be modified in the field, as appropriate, based on the professional judgment of the field team leader (FTL). All modifications will be documented in the field logbook or on the field sampling data sheets. Soil samples collocated with the plant tissue samples (composited from each corner and the center of the 100×100 -m [110×110 -yd] grid) also will be collected.

B-2.1.2 Mammal Sampling Procedure for Analytical Sampling

One small mammal species, the deer mouse (*Peromyscus maniculatus*), representing the major links between primary and secondary consumers and higher predators, will be collected for tissue analyses. The deer mouse is a primary prey item for both secondary and tertiary consumers. This species is commonly used to represent several important linkages in the food chain and is the primary choice because it is omnivorous, widespread, and relatively easy to collect.

Collection of animal samples will be performed in accordance with applicable sections of TPR-145 and the following information. Deer mice will be collected for tissue analysis. Typically, it will be necessary to collect several deer mice for each analysis to obtain the 60 g of tissue required. Deer mice will be composited to obtain the required tissue amounts. Compositing will not include segregation of small mammals by sex or age but will be limited to the single species. Small mammal species—other than deer mice—will be weighed, photographed, have other life history or details recorded in the field logbook, and released.

The deer mouse samples will not be washed before homogenization. The intent of this sample preparation is to evaluate the body parts that a predator is most likely to consume. By incorporating all unwashed biotic tissue, all available contaminants in each sample will be assessed; however, not all analytes are necessarily bioavailable.

The same trapping design (see Section B-3.1.3) used to evaluate small mammal population/community data will be used to collect deer mouse tissue samples for analytical assessment. Ten trapping locations or sample plots will be used in each grid. Each sample plot will require a 2- to 3-week trapping period and will consist of 100 traps placed along 10 parallel transect lines (10 traps on each). Each transect will follow a roughly straight line 100 m (110 yd) long. An example of the transect design is shown in Figure B-1.

At each plot, traps will be opened Monday afternoon and left open (weather permitting) 3 nights (Monday through Wednesday night), closed 4 nights (Thursday through Sunday), and then reopened an additional 3 nights (Monday through Wednesday night). If the weather becomes too hot, it may be necessary to close traps during the day to minimize mortality of diurnal species. Once an animal is

trapped, a uniquely numbered ear tag will be attached. The ear tag correlates with the trap location, genus, species, collector's initials, and date recorded in a field logbook. The animal should be emptied into a plastic bag. It should be sexed, aged (adult/juvenile), weighed, and identified to its species if possible. A ruler should be used to measure the head-body length, ear (from skull to tip), tail, and right hind foot to the nearest millimeter. The animal should then be returned for release to the location it was trapped. All information should be recorded on the data sheet.

Tissues will be collected for chemical and radiological analysis, genetics, and histopathology. On the last day of the population surveys, at least three deer mice in each grid will be retained as a single composite sample. Animals to be sacrificed for contaminant analysis will be dispatched in the field. After dispatch, each carcass will be weighed and placed in another clean plastic bag. The amount of sample material in the composite sample will be determined by summing the weights of the individual specimens from each location. Processing should take place as soon as possible after checking traps to reduce potential degradation of the specimen. Samples will be placed on ice for transport to the processing center.

Portions of each animal's liver and kidney will be collected for weight and histopathology. A ventral incision will be made with a clean scalpel blade. The liver and kidney will be removed and weighed to the nearest 0.01 g. Small sections of the liver and kidney will be sliced and placed in a 10% buffered formalin. This solution is potentially carcinogenic and should be handled with caution that is detailed on the respective material safety data sheets. The jar will be labeled with appropriate sample information (i.e., time, date, and sample identification number). Small sections of maternal and fetal tissue will be removed from female mice. The carcasses will be placed in a sealable plastic bag and placed inside another bag with the sample labeled. Chain-of-custody forms will be filled out.

Tissue samples for residue analysis should be frozen and shipped on Blue Ice (or equivalent) to the laboratory. Dry ice can cause serious skin burns if handled incorrectly. Gloves should be worn when handling dry ice.

A single voucher specimen will be photographed but will not be analyzed for contaminants. An experienced wildlife biologist will examine the voucher specimen to verify genus and species.

B-2.2 Soil Analytical Characterization

Soil samples will be collected from the surface 0 to 5 cm (0 to 2 in.) and subsurface 5 to 61 cm (2 to 24 in.) or bedrock (i.e., limited to two sampling intervals) and will consist of composites from locations within the sampling plot designs that correspond to plants from which vegetation samples are collected.

Before sampling, it is important to calculate the total volume of sample material that will be needed from each increment sample location to ensure that the volume required for each analysis is available to completely fill each sample container. The analysis-specific volumes are specified in the Attachment A field guidance forms. Sampling locations specified will be identified and marked using surveying stakes, lath, or flags. The soil will be evaluated for contamination concentrations.

B-2.2.1 Surface Soil Material

Composite surface material samples will comprise five increment subsamples collected from each of the corners and center point of a 100-m (110-yd) square. All or a portion of the increment samples will be mixed together to create a composite sample representative of average constituent concentrations within the area to be investigated. For a given composite sample, the volume of each increment sample

must be the same and must equal 1/n of the required composite sample volume, where n equals the number of increment samples making up the composite sample.

Surface material samples will be collected as follows:

- 1. At each subsample location, an area approximately 61 cm (24 in.) in diameter is cleared of surface vegetation, nondecomposed plant litter, and debris.
- 2. A decontaminated stainless-steel spoon or hand auger is used to collect surface material to a depth of 5 cm (2 in.). A stainless-steel pick can be used as needed to loosen the soil. To the extent possible, gravel-size or larger particles and debris are eliminated, based on visual observation.
- 3. The material is described visually, and observations are recorded on the soil sample field data sheet.
- 4. The increment sample is sieved through a No. 10 mesh, and the fine fraction is placed into a decontaminated stainless-steel mixing bowl and then thoroughly mixed.
- 5. For composite samples, Steps 1 through 4 are repeated at each increment sample location for that composite sample, adding each successive increment sample to the mixing bowl.
- 6. The sample material is thoroughly mixed in the stainless-steel bowl using a decontaminated stainless-steel spoon. To homogenize the sample, it is divided into four quarters and each quarter is mixed, then the four quarters are combined and the entire sample is mixed. The mixture is placed into the appropriate laboratory-supplied sample containers.
- 7. The containers are labeled and handled as required. Soil subsample location descriptions and collection information will be documented in the logbook in accordance with Management Control Procedure (MCP) -1194, "Logbook Practices for ER and D&D&D Projects."

B-2.2.2 Subsurface Soil Material

Subsurface material samples will be collected as composite samples. Before sampling, it is important to calculate the total volume of collected sample material at each increment sample location to ensure that the volume required for each analysis is available to completely fill each sample container. The analysis-specific volumes are specified in the Appendix A field guidance forms. Specified sampling locations will be identified and marked using surveying stakes, lath, or flags.

Composite surface material samples will comprise five increment subsamples collected from each of the corners and center point of a 100-m (110-yd) square. All or a portion of the increment samples will be mixed together to create a composite sample representative of average constituent concentrations within the area to be investigated. For a given composite sample, the volume of each increment sample must be the same and must equal 1/n of the required composite sample volume, where n equals the number of increment samples making up the composite sample.

Subsurface material samples are collected as follows:

1. At each sample location, an area approximately 61 cm (24 in.) in diameter is cleared of surface vegetation (nondecomposed plant litter) and debris.

- 2. A decontaminated stainless-steel spoon or hand auger is used to collect subsurface material from a depth of 5 cm (2 in.) to no more than 61 cm (24 in.) below ground surface. A stainless-steel pick can be used as needed to loosen the soil. To the extent possible, gravel-size or larger particles and debris are eliminated based on visual observation. Depth must be recorded for each soil core collected.
- 3. The material is visually described, and observations are recorded on the soil sample field data sheet.
- 4. The increment sample is sieved through a No. 10 mesh, and the fine fraction is placed into a decontaminated stainless-steel mixing bowl and then thoroughly mixed.
- 5. For composite samples, Steps 1 through 4 are repeated at each increment sample location for that composite sample, adding each successive increment sample to the mixing bowl.
- 6. The sample material is thoroughly mixed in the stainless-steel bowl using a decontaminated stainless-steel spoon. To homogenize the sample, it is divided into four quarters and each quarter is mixed, then the four quarters are combined and the entire sample is mixed. The mixture is placed into the appropriate laboratory-supplied sample containers.
- 7. The containers are labeled and handled as required. Soil subsample location descriptions and collection information will be documented in the logbook in accordance with MCP-1194.

The center of the sample grid location will be surveyed using a Global Positioning System unit.

B-2.3 Soil Nutrient and Physical Characterization

Soil samples for soil nutrient and physical characterization will be collected at the same locations as soil samples for contaminant analysis. Each composite sample will be collected as follows:

- Soil sampling sites will be collocated with chemical and radiological soil samples.
- After collection of the chemical analysis samples (described above), appropriate amounts of homogenized soil will be placed into the shipping containers for analysis. Approximately 500 g will be placed into a sealable plastic bag for soil nutrient and physical characterization.
- The containers will be labeled and handled as specified in the field sampling plan (FSP).

These procedures can be modified in the field, as appropriate, based on the professional judgment of the FTL. All modifications will be documented in the field logbook or on the field sampling data sheets.

B-3. EFFECTS SAMPLING

B-3.1 Population/Community Data

Ecological systems such as populations or communities are usually quite large and complex. These systems must be described and quantified to compare them with one another or assess changes in them. Several ecological variables can be measured (e.g., density, frequency, coverage, and biomass) to describe populations and communities. These measurements are used to characterize aspects of

populations and communities such as presence/absence, population density, population distribution, species diversity, and productivity (biomass).

B-3.1.1 Vegetation

Fifty Daubenmire quadrats will be collected at each of the 10 AOC plots. Transects will be located between each of the 10 trapping lines (see Figure B-1) in each 100×100 -m plots. Each transect line will have five quadrat locations spaced approximately 2 m (6 ft) apart. These locations will be selected by striding 20 to 25 paces between quadrats starting at the edge of the 100×100 -m plot. The quadrat frame will be placed with the left side of the short end of the frame at the edge of the right foot. A 1×3 -m (1.1×3.3 -yd) quadrat will be used to estimate percent ground cover. As the quadrat frame is placed along the tape at the specified intervals, the canopy coverage of each plant species will be estimated. In addition, the data will be recorded by quadrat, species, and cover class. Canopy coverage can be estimated, as follows, for both perennial and annual plant species:

- 1. The quadrat frame is observed directly from above, and the cover class for all individuals of a plant species in the quadrat is estimated as a unit. All other kinds of plants are ignored as each plant species is considered separately.
- 2. A line drawn about the leaf tips of the undisturbed canopies (ignoring inflorescence) is imagined, and these polygonal images are projected onto the ground. This projection is considered "canopy coverage." The classes that the canopy coverage of the species falls into can be determined (see Table B-1).
- 3. Canopies extending over the quadrat are estimated even if the plants are not rooted in the quadrat.
- 4. The data are collected during a period of maximum growth for key species.
- 5. For tiny annuals, it is helpful to estimate the number of individuals that would be required to fill 5% of the frame. A quick estimate of individuals in each frame will then provide an estimate as to whether the aggregate coverage falls in Class 1 or 2, etc.
- 6. Overlapping canopy cover is included in the cover estimates by species; therefore, total cover might exceed 100%. Total cover might not reflect actual ground cover.

Table B-1. Plant cover classes.

Coverage Class	Range of Coverage (%)	Midpoint of Range (%)
1	0 to 5	2.5
2	6 to 25	15.0
3	26 to 50	37.5
4	51 to 75	62.5
5	76 to 95	85.0
6	95 to 100	97.5

While using this method, it is important to keep track of the growth form of each species so that comparisons of grass vs. forb vs. shrub can be made. In addition, an estimation of the cover of bare ground and rocks will provide additional characterization data. While conducting this survey, it is

important to remember to record total cover for each quadrat, because this might differ from the sum of the cover values for individual species (due to plant canopy overlap). The surveyor should have a cover category for each quadrat among all identifiable species, mosses (if any), bare ground, rocks, and total cover.

Within each quadrat, the shrub height will be measured by species. To measure shrub height, one person will hold a telescoping rod or other measurement device in the center of a shrub while the other person records the height. If no shrub is present within the plot, the closest shrub(s) to the quadrat of each of the dominant species will be measured.

Once the surveys are complete, the species cover can be estimated by multiplying the number of times a class is recorded by the midpoint of that cover class, adding the results for each class, and calculating an average by dividing by the total number of quadrats sampled. Data are usually collected from many quadrats located along a transect, so that the transect is the sample unit. Therefore, data must be collected from several transects to determine the sample's precision for statistical analysis of cover data.

This method recognizes the difficulty in accurately assigning an exact percent cover value to each quadrat, because even highly experienced workers are unlikely to visually estimate closer than about 5% cover. Assigning broad cover classes provides an equally accurate result as long as the data follow a normal distribution around the midpoint within each class. The narrower upper and lower classes of the Daubenmire scale protect against skewed data in extremely sparse or dense vegetation.

Ranking the data into broad classes is also a relatively rapid procedure because observers are not required to spend as much time contemplating quadrat cover to the nearest percent. In fact, rapid evaluation of each quadrat is the key to success with this approach, since a large sample is less sensitive to the occasional incorrect ranking.

B-3.1.2 Avian

The avian wildlife on designated study areas at the INL will be monitored with point counts and nest searches. Avian point counts will be conducted to assess species occurrence and relative abundance in each study area. Point counts have been used throughout North America for long-term bird monitoring programs such as the Breeding Bird Survey (BBS). The BBS is a roadside route survey of avifauna designed to monitor abundance and distribution of birds in both the U.S. and southern Canada. It began in the eastern U.S. in 1966 but is now nationwide in scope (Bystrak 1981; Robbins et al. 1986). Since 1985, official BBS and modified "mini-routes" have been surveyed at the INL (Belthoff and Ellsworth 1999). Nest searches will be used to evaluate the feasibility of harvesting eggs for toxicology research.

Each area of concern (i.e., each WAG) will be divided into 10 randomly chosen 100 m² plots according to the small mammal trapping protocol. These plots will be used as a reference for designing walking or driving routes through each area. Routes will be designed to survey an area similar to that covered by the mammal plots, and points will be located near or in the plots as often as possible. Points will be located at least 400 m from the nearest neighboring point. Each point will be named, flagged, and marked using a global positioning system (GPS). Each route consists of 10 point count locations. The route for the reference area was established in 2004 and will remain the same in subsequent years.

Breeding Bird Surveys are conducted during the peak of the nesting season, primarily in June, although surveys in desert regions and some southern states (where the breeding season begins earlier), are conducted in May (http://www.mbr-pwrc.usgs.gov/bbs/genintro.html). Similar to the surveys already established on the INL, the LTEM project surveys will be conducted from mid June to early July. Surveys

will be performed only when weather conditions are satisfactory as prescribed by the BBS protocol. Temperature, wind speed, and cloud cover at the start and end of each survey will be recorded in an entry form as presented in Attachment A.

Each point along a route will be the site of one 3-minute, unlimited radius point count. At each point an observer will count all the individuals seen or heard within the allotted time period. Counting the same individual twice should be avoided even if encountered at different count locations. Surveys begin approximately one-half hour before sunrise and continue until three replicates are completed, with at least 45 minutes elapsing between the start of one replicate and the start of the next. This avoids potential bias that disturbance may cause in subsequent replicates. Surveys will not be conducted during inclement weather, which includes any amount of precipitation, wind exceeding 12 mph, or other conditions that interfere with detecting birds by sight or sound.

Nest searches will also be conducted within the designated mammal plots. Surveyors will systematically walk through each grid with drag lines, or by visual inspection, and flag the location of any nests that were found and record the species and other pertinent information about the nest site. Nests will be digitally photographed.

B-3.1.3 Small Mammals

Small mammals will be evaluated by using live trapping methods. The 10 sample plots established for biota and soil analytical sampling will be used to assess the small mammal population/community data in the sampling area. Each sample plot will require a 2- to 3-week trapping period and will consist of 100 traps placed along 10 transect lines (10 traps on each) in a line grid formation. Each of the transects will approximately follow a 100-m-long (110-yd-long) straight line. An example of the transect design is shown in Figure B-1.

Traps will be left open four nights, closed three nights, and then reopened an additional four nights. Once an animal is trapped, a uniquely numbered ear tag will be attached. The ear tag will correlate with the trap location, genus, species, collector's initials, and date recorded in a field logbook. The animal should be emptied into a plastic bag. It should be sexed, aged (adult/juvenile), weighed, and identified to its species if possible. A ruler should be used to measure the head-body length, ear (from skull to tip), tail, and right hind foot to the nearest millimeter. The animal should then be released to the original location from where it was trapped. All information should be recorded on the data sheet.

The mark-and-recapture method will be used in estimating population densities. This method involves several steps:

- 1. Trapping and marking some individuals of a population
- 2. Releasing the known number of marked individuals back into the population from which they were captured
- 3. Trapping some individuals of the population after the marked individuals have had a chance to redistribute themselves into the population
- 4. Estimating the total population size by a series of computations that are based on the ratio of marked to unmarked individuals in the recapture attempt.

Generally speaking, if the population is large, the marked individuals will become diluted within the population and only a few of the marked individuals would be expected to appear in the second sample. If assumptions about the sampling and animals' distribution are correct, then the proportion of marked individuals in the second sample would be the same as the entire population.

Like all estimation procedures, a number of assumptions must be met to validly use this method:

- The two samples taken from the population must be random samples (i.e., all individuals in the population have an equal and independent chance of being captured during the time of sampling).
- There is no change in the ratio of marked to unmarked animals, meaning that from initial capture to recapture, there must be no significant addition of unmarked animals to the population through births or immigration.
- The population losses from mortality and emigration must remove the same proportion of marked and unmarked individuals.
- The marking of individuals does not affect their mortality.
- Individuals do not lose marks.

The Peterson-Lincoln Index, the simplest method for determining the population size, will be used. The total population can be estimated as follows:

- Assume the total estimated population size contains N individuals.
- Sample M individuals from this population, mark these animals, and return them to the population.
- Sample a second set of n individuals from the population; this sample contains recaptured animals (i.e., individuals captured and marked in the first sampling).
- Estimate the population size, N, by the following equation.

$$N = Mn/R \tag{B-1}$$

Equation (B-1) might overestimate the population size (i.e., it is biased) when samples are relatively small. No is a nearly unbiased estimate of population size if the number of recaptured animals, R, is at least eight. Using Equation B-2 can reduce this bias:

$$Nc = \frac{(M+1)(n+1)-1}{R+1}$$
(B-2)

The approximate variance, s², of this estimate is in Equation B-3 below:

$$s^{2} = \frac{(M+1)(n+1)(M-R)(n-R)}{(R+1)^{2}(R+2)}$$
(B-3)

With the standard deviation, s, 95% and 99% confidence limits on the population estimate are given by Equations B-4 and B-5 below:

$$N (or NC) + 1.96(s)(95\% confidence limits)$$
 (B-4)

and

$$N \text{ (or NC)} + 2.58(s)(99\% \text{ confidence limits)}.$$
 (B-5)

B-3.1.4 Reptiles

B-3.1.4.1 Introduction. Limited studies have been performed at the INL on reptiles such as the sagebrush lizard and horned lizard. These species, since they presumably have a smaller home range, may be excellent candidates for future monitoring at the sites of concern. The reptiles at the INL appear to have a very patchy distribution and are highly dependent on weather conditions for their metabolic activities. This means that the catchability of reptiles is greatly affected by the sampling design as well as the weather during the sampling period.

Common methods used to sample reptilian populations are:

- Intensive and systematic searching
- Transects
- Pitfall and cover trapping.

The first method basically means that the herpetologist visits every habitat and accurately searches for reptiles. Microhabitats potentially occupied by reptiles include a variety of shelters. Reptiles can be found by overturning rocks, logs, and other "cover" elements, such as sheets of scrap metal, plastic, etc. These sheets may provide a very attractive microhabitat to many reptiles in the spring and early summer since these sheets accumulate warmth. Overturned rocks and twigs should always be replaced in their original position to avoid modifying the habitat. Data on the relative abundance of a given species may be standardized with the searching effort, but these results are highly biased by the experience and ability of the observer.

Transect sampling is usually used for terrestrial species. The investigator first selects the transect length and width and the search effort. Many transects are at least 50 m × 4 m. People commonly use search effort to "define" the length of the transect, so that transects from different locations can be compared even if we don't know the exact transect length. Search effort is defined as number of hour*person (i.e., one person searching during one hour will be 1 hour*person, while 2 persons searching during one hour will be 2 hour*person).

When sampling reptiles, it is usually better to sample favorable habitats at the best time possible (i.e., when reptiles are starting to bask). This usually happens in the early to mid morning depending on the latitude and the weather. Cloudy days are usually better than fully sunny days to observe reptiles. Basking reptiles can be counted along a transect of a known length, or their number may be related to the time spent looking for them. The observer should walk slowly along the transect.

In an area of good habitat, drift fencing may significantly increase the trapping success. The fence should be at least 40 cm high and can be made of a variety of materials (e.g., sheet metal, hard plastic, or plastic sheet). Depending on the site, fences can encircle a hibernaculum. Pitfall traps, such as plastic

buckets, are dug into the ground on both sides of the fence. To avoid water filling the buckets, the traps should have several holes on the bottom. It may also be useful to leave a plastic cover for the buckets between two sections of fence so that when sampling is not possible or desirable, the cover can be removed from the fence and placed over the bucket. Depending on the density of animals, buckets should be placed at a distance of 5 to 25 m along the fence. Buckets must be checked at least once a day, preferably twice. Ideally, animals should be removed from the buckets early in the morning and late in the evening. A variety of amphibian predators can be attracted by the traps; amphibians themselves can also suffer from dessication or freezing. Other animals, especially rodents, can also fall into the traps and are much more vulnerable to die because of their faster metabolism.

Some disadvantages of drift fencing are its high cost and the labor requirements to install fences and control the traps. Also drift fencing can attract predators or be destroyed by large vertebrates or by people. A reduced version of the drift fencing explained above is to cross two short (10 to 50-m) sections of fences and install buckets inside each of the four squares defined by the fences. This design has been used in temperate and tropical forests to sample leaf-litter amphibians.

Pitfall traps used to sample reptiles vary in size according to the target species. Fences are not used to capture reptiles (many reptiles will easily climb them); instead, pitfall traps are placed very close to potential microhabitats. Pitfall traps must be checked frequently for amphibians.

B-3.1.4.2 *Idaho National Laboratory Approach.* Several methods will be evaluated in 2005 and 2006 to determine which method or combination of methods will provide the best approach for monitoring the sagebrush and horned lizard at the sites. This includes a combination of observation study, track plates, and intensive surveys.

The pitfall traps that are located on the two plots at the reference area during 2004 will be reopened and data will be collected for the two weeks during small mammal trapping at these areas. From the 2004 data, pitfalls do not appear to be highly successful in these areas. This is most likely because they are not in areas of high density for these species. However, the data collected this year will be combined and compared to 2004 information.

B-3.1.4.3 Observational Study, Transects, and Track-Plates. At each plot, each team member will watch for and record reptile activity while checking traps during the morning and afternoon. If possible, when checking the traps, technicians should walk facing north or south because it is easier to detect lizards if the sun is to the right or left of the person.

In the morning, when a reptile is observed, the recorders will write down the plot number, the time each plot is surveyed, the air temperature, and what species were seen. Also surrounding habitat and weather (cloud cover) will be recorded.

On the last day of the first week of small mammal trapping, sheets of aluminum or other types of material to provide an attractive microhabitat will be located across a selected number of plots at even intervals (depending on ability to place a sheet). Ten sheets of approximately 3×3 ft will be used.

At these plots, tracking plates and possibly scent stations will be used for animal and reptile detection by luring them to a scent and recording their footprints in a tracking medium. Scent stations consist of 1 × 1-m aluminum plates, heavily smoked with flame, as the tracking surface (Barrett 1983). The scent station attractant, cat food, or predator survey discs will be placed in a slightly elevated position at the center of the station. Stations will be checked for visitation early each morning and tracks will be cleared when necessary. Tracks can be lifted from the plates by lightly pressing with a wide piece of transparent tape. Track outlines will be identified and placed in notebooks for future reference. Track

plates should be cleaned and the tracking medium replaced when rain, heat, or signs disturb the carbon-coated surface.

B-3.1.4.4 Intensive and Systematic Searching. Intensive and systematic searching will be performed on a selected number of plots at each area of concern. The plots selected will be based on the observational data collected during the mammal trapping.

If captured, collection data recorded (see Attachment A) includes; sex, weight (g), snout-vent length (mm), and tail length (mm). A unique number will be written with permanent marker on the ventral surface to identify the individual. Painting the side or leg with a noticeable color of fingernail polish changing day-to-day may also be used to help identify an already-captured animal without recapturing. If not captured, the species and location will be noted.

Note the following when performing intensive searching of terrestrial plots:

- The use of gardening gloves is recommended to avoid bites from ants and other invertebrates.
- When sampling areas that are known habitats of venomous snakes, care should be taken to avoid getting bitten. Hand-held rakes or small sticks can be used to search the leaf litter and when turning logs and rocks. Always turn the log or rock toward you, so that if an aggressive snake is present, your feet are somewhat protected by the log or the rock.

B-3.2 Earthworm and Plant Bioassay Soil Samples

Bioassay soil samples will be collected at each plot. Each composite sample will be collected as follows:

- Soil will be taken evenly from 0 to 30 cm. A composite sample of 4 gals will be collected from five locations at each plot. The five locations will be from the center and four corners of the plot.
- Containers will be labeled with the date, location, and other appropriate information and shipped on ice to the bioassay laboratory for processing.

These procedures can be modified in the field, as appropriate, based on the professional judgment of the FTL. All modifications will be documented in the field logbook or on the field sampling data sheets.

B-3.3 Soil Fauna^a

B-3.3.1.1 The Animals to be Enumerated. Microarthropods comprise two soil fauna groups: (1) Collembola, also known as springtails, and (2) Acari, also known as mites. The microarthropods are typically the most abundant soil animals in surface layers, especially in association with litter inputs from plants. The size range for microarthropods is 0.1–2.0 mm. Many of these animals are fungus feeders, but many are also predators on each other and on nematodes and flies' eggs. There is a great diversity of species of microarthropods in soil, and considerable time is needed to identify each specimen to species. The approach taken is usually to divide the animals into major subdivision groups within the Collembola

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a. Procedure of the laboratory of Dr. Terence McGonigle (Brandon University, Manitoba, Canada) for the determination of soil fauna of microarthropods in a field sample of surface soil with plant litter.

and Acari for counting purposes. The animals occur in the field densities in the range 1,000–50,000 per square m, although the lower part of this range is expected for dry environments.

B-3.3.1.2 Collection of Field Samples. Samples are to be collected as undisturbed soil discs. One sample will be taken from each plot at each area of concern. Each plastic disc circular is 77 mm in internal diameter and 88 mm in external diameter, has a 4-mm depth, and a 45-degree bevel on one outside edge to permit entry into the soil. Each disc is driven flush into the soil using a hammer and wooden board placed above. The disc is then removed intact with litter using a hand trowel and placed upright on a flat surface for immediate transportation to the laboratory.

B-3.3.1.3 Extraction of Fauna from Field Samples. The fauna are subjected to an active extraction for removal from soil. The active extraction can be summarized as applying heat and light above the sample so that the animals walk out the bottom of the disc, where they are collected. The active extraction takes 3–7 days to complete, but it has the advantage of recovering the animals in good condition and mostly free of soil and other debris. Physical separation of animals from field samples by use of high-density fluids is not preferred because animals are typically recovered in poor condition. Full details of available extraction methods are given in "Soil Invertebrates" (Coleman et al. 1999) and "Standard Soil Methods for Long-Term Ecological Research" (Robertson et. al 1999).

Discs returned to the laboratory will be moistened by adding 20 ml water to the surface using a pipette and placed upper surface down into extraction units established in the laboratory. Twelve extraction units can run in the laboratory simultaneously so that three field collections will be processed in separate extraction runs to make 36 samples in all. Each extraction unit has a fiberglass screen nylon mesh supported on a coarse wire grid above a funnel that collects to a plastic vial. A screw-fit lid with an upper airway covers the disc sample to delay moisture loss. A single light with a 60-watt bulb is placed immediately above each disc for up to 7 days, and the fauna are collected in 1-cm depth of 70% ethanol in the vial. Each vial is checked and emptied daily until fauna no longer appear. The total fauna extracted for a given disc is then pooled and stored in 70% ethanol.

B-3.3.1.4 Sorting and Enumeration of Extracted Fauna. All subsequent laboratory work will involve use of the stereoscopic microscope in the laboratory by a knowable expert in this field. Each sample is sorted, using a Pasteur pipette, into major groups within the Collembola and Acari, and the number of animals for each group will be counted. The sorted samples are stored in 70% ethanol in separate glass vials, one for each major group for a disc. The major groups recovered are expected to be as follows:

Collembola ^a	Acari
(All are fungivores)	Mesostigmata (predatory)
Podurids	Oribatids adult (fungivores)
Entomobryids	Oribatids juvenile (fungivores)
Neelids	Astigmata (fungivores)
Symphyla	Prostigmata (most are fungivores, predators can be separated)

a. Collembola groups are further subdivided into epigeous (pigmented, seeing) and hypogeous (non-pigmented, blind) life forms.

B-3.3.1.5 Data Report. The data will be returned as a report summarizing the counts of each major group, as described above, with tallies taken separately for each group for each disc. The report will be submitted along with the fauna samples. Spent soil and litter contents of each disc will be returned to the site.

B-3.4 Histopathology and Body and Organ Weight

Tissues will be collected from small mammals for chemical and radiological analysis, genetics, and histopathology. On the last day of small-mammal population surveys (see Section B-3.1.3), at least three deer mice in each sampling plot will be retained as a single composite sample. Deer mice will be humanely harvested by cervical dislocation or asphyxiation with carbon dioxide gas before transport to the laboratory. Animals should be removed from traps one at a time, so that specimens are not misidentified. Processing should take place as soon as possible after trap checks to reduce potential degradation of the specimen. The deer mice will be weighed in the laboratory to the nearest 0.01 g.

A ventral incision will be made with a clean scalpel blade. The liver and kidney will be weighed to the nearest 0.01 g. Small slices of each will be placed in 10% buffered formalin and the rest will be returned to the carcass. This solution is potentially carcinogenic and should be handled with caution, as detailed on the material safety data sheet. The jar will be labeled with appropriate sample information (time, date, sample identification number, and ear tag number).

The carcasses forming the single composite sample will be placed in a sealable plastic bag, placed inside another bag, and then labeled for contaminant analysis. Chain-of-custody forms will be filled out.

The removal of the kidney and liver may slightly reduce apparent concentrations. Estimated loss in concentration is as shown in Equation (B-6):

$$mg/kg WB * kg WB + mg/kg L * kg L + mg/kg k * kg k$$
 (B-6)

where:

mg/kg WB = concentration in whole body

mg/kg L = concentration in liver (estimated)

mg/kg k = concentration in kidney (estimated).

A bioaccumulation factor from the literature will be used to estimate the fraction lost to histopathology. Although the bioaccumulation factor introduces uncertainty into the assessment, the liver and kidney tend to concentrate metals and might exhibit cellular changes for evaluation of effects from exposure. If effects are determined to be present, a selected study will be performed to further characterize this problem, or the sampling approach will be modified appropriately.

B-4. AQUATIC ECOSYSTEM CHARACTERIZATION

Chilly Slough was selected as the aquatic reference area.

B-4.1 Sediment and Surface Water Analytical Sampling

Sediment and surface water samples will be obtained from the reference area and from the waste ponds at the TAN Technical Support Facility (TSF) -07 if water is available. The data will be used to predict health effects and exposure in aquatic receptors. Five grab samples of each medium will be collected from a 10-m² (108-ft²) grid surrounding the pond. If water is unavailable, the TAN TSF-07 disposal pond will be treated as a terrestrial sampling area.

B-4.2 Biota Analytical and Effects Sampling

If appropriate aquatic receptors are identified and present, they will be collected and identified to the lowest taxonomic level possible. Sixty grams is required for all analytical work. Five samples will be collected from the pond.

Attachment A Survey Forms and Data Sheets

Date	
Weather	
Conditions _	
Cloud cover	

Reptile Data Sheet

location	plot #	nearest trap #	species	time (24 hrs)	air temp (degree C)	sex	weight (g)	SVL (mm)	tail (mm)	habitat/comments

Page	Number	
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Date Weather Conditions Cloud cover		Reptile Data Sheet											

Name Date				_	Daubenmire Plot Form								Location Plot Number																	
DaubPlot #																														
Species	1/4	1/2	1	1/4	1/2	1	1/4	1/2	1	1/4	1/2	1	1/4	1/2	1	1/4	1/2	1	1/4	1/2	1	1/4	1/2	1	1/4	1/2	1	1/4	1/2	1
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Daubenmire	DataSheet
Checker	

Recorder's Name	
Trapper's Name	

Small Mammal Data Sheet

Date	
Location	
Plot #	

	Trap#	Species	Tag #	Body (mm)	Tail (nm)	Foot (nm)	Ear (nm)	Sex	Mass g)	Recapture	Dead?	Other
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Checker signoff	

Nest Count Data Form

Date	Location	Nest ID	Species	# of eggs	# of young	Nest Site Description	Comments

Observer: Date:		Avian Po	TemperatureWindCloud cover			
Location	Time	Species	Tally			Notes
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